

1 **Bio-economic Modeling for Swine Genomic Selection Index Design and Multi-**
2 **Generational Response Simulation**

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12 **Abstract**

13 The selection index is related to the efficiency and direction of genetic improvement for
14 livestock. Nevertheless, specialized software that employs multiple bio-economic models to
15 design and compute the selection index and predict the multi-generational selection response is
16 still lacking. In this study, to meet the needs of pig breeding programs, visualized multivariate
17 model-based software for pig selection index design and computation, CSIP, was developed to
18 rapidly construct a selection index that is applicable to different pig breeding systems, such as
19 farrow-to-finish herds, single pure-line, two-way cross, and three-way cross breeding programs,
20 and to predict multi-generational selection response. The main advantages of CSIP include full-
21 process visualization for the design of a selection index, which increases transparency and user
22 comprehension; incorporation of multi-generational selection response, which facilitates the
23 proactive mitigation of antagonistic trait response and suboptimal genetic gain; integration of
24 gene flow and economic weight sensitivity analysis, which increases robustness and prevents
25 errors in economic weight calculation; and an accessible, workflow-based interface that
26 supports flexible calculation strategies to accommodate diverse practical breeding scenarios.
27 These features position CSIP as an innovative tool for increasing the precision, efficiency, and
28 economic viability of pig breeding programs.

29 **Keywords:** Gene flow; Multi-generational selection response; Bio-economic model;
30 Selection index

31 **Introduction**

32 In pig breeding, the simultaneous improvement of multiple traits is often required to meet
33 production and economic goals. To address this requirement, the selection index method has
34 been widely adopted, as it integrates information from various traits into a single numerical
35 value by combining genetic and economic parameters [1,2]. A selection index is typically a
36 linear combination of estimated breeding values (EBVs) and their corresponding economic
37 weights, where the economic weights reflect the relative economic importance of each trait [3-
38 5]. The ultimate goal of animal breeding is to maximize the overall selection response in
39 alignment with a defined economic objective [6,7]. Therefore, accurately determining the
40 economic weight of each trait is central to the construction of an effective selection index.

41 The estimation of economic weights for different traits in the pig selection index has been
42 explored extensively since the concept of the selection index was proposed [8-15]. The simple
43 profit–function approach, which uses a simple function to calculate the partial derivative of
44 traits to obtain economic weights [10,16,17], is often inadequate for complex pig breeding
45 systems; subsequently, bio-economic models can be established to derive economic weights
46 [8,9,11,18-20]. At the initial stage of using the selection index, marginal economic values
47 (MEVs) were regarded as the economic weights for different traits [1,9]. However, real pig
48 production systems involve distinct population structures (nucleus, multiplier, and commercial
49 herds), time delays in trait expression between parents and offspring, and varying gene
50 expression across breeds, rendering the use of simple MEVs as economic weights insufficient
51 [21]. Therefore, economic weights in bio-economic models can be calculated as the product of
52 a trait’s MEVs and its corresponding number of discounted gene expressions (NDEs) to capture

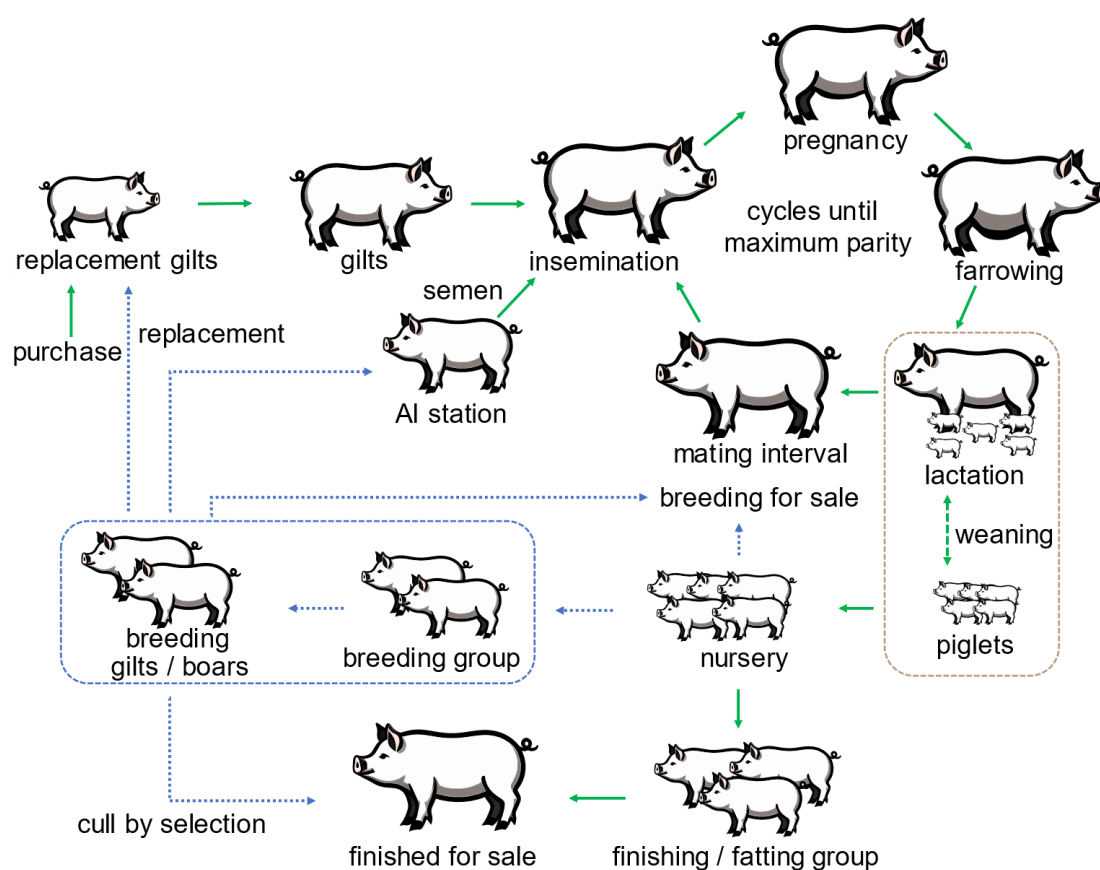
53 both the trait's economic impact and its transmission dynamics across generations [22,23]. The
54 NDEs reflects the accumulation of selected parental genes in progeny across generations and
55 production tiers, with trait-expressing genes discounted back to the year of parental selection
56 using time-adjusted discount rates [23,24]. For compute NDEs, a gene flow method [24,25],
57 which has been successfully applied to optimize pig breeding programs [26-28], was applied to
58 quantify gene expression. To expand the scenarios for the use of bio-economic models, Wolfová
59 *et al.* [29] developed a flexible bio-economic model to calculate the MEVs of traits in diverse
60 pig breeding systems, and subsequently, a computation program was developed to calculate
61 economic weights [23].

62 After a selection index is formulated, it is essential for breeders to evaluate whether the index
63 maximizes the multi-generational selection response [30]. This evaluation not only verifies the
64 alignment between the index and the breeding objectives but also facilitates the refinement of
65 breeding strategies and the optimization of index design. However, studies of the integration of
66 multi-generational selection response prediction with the formulation of selection indices are
67 still lacking. Moreover, there is currently no software that supports the full workflow—from
68 index formulation to multi-generational response prediction and visualization—which limits
69 the effective and practical application of the selection index in pig breeding.

70 The goal of this study is thus to develop an intuitive, computationally efficient, and fully
71 visualized tool to support the formulation of a pig selection index in diverse breeding scenarios
72 while enabling the prediction of multi-generational selection response. To achieve this goal, we
73 constructed bio-economic models for farrow-to-finish (commercial) herds and different pig
74 breeding systems (including pure-line, two-way cross, and three-way cross breeding systems)

75 to derive MEVs; integrated the gene flow approach into distinct pig breeding systems to
76 calculate NDEs and the corresponding economic weights; incorporated predictions of multi-
77 generational selection response to validate the rationality of the formulated index; and
78 streamlined the index formulation workflow for user accessibility and flexibility in its
79 implementation. CSIP features built-in sensitivity analysis for economic weights, standardized
80 index computations, and comprehensive visualization tools to meet both research and industry
81 demands.

82



83

84 **Figure 1.** Basic framework of the bio-economic models for farrow-to-finish and pure-line herds.

85 The diagram illustrates the integrated production system, encompassing the life-history of

86 breeding sows (reproduction and growth) and the complete rearing process of the offspring.

87 The part connected by the blue box and the dotted line is included only in the pure-line model,
88 and the remaining parts are contained in both models.

89

90 **Methods**

91 In this study, several bio-economic models were established to derive economic values for
92 different pig traits. Among these models, the commercial farrow-to-finish herd model was
93 adapted from De Vries [31]. In addition, we modeled the standard pure-line, two-way cross,
94 and three-way cross breeding systems. These structures were selected as they represent the most
95 widely adopted production systems in the global swine industry, where artificial insemination
96 (AI) is the standard practice. Within the CSIP, these models were parameterized to construct
97 deterministic profit functions, integrating twenty-six specific traits to derive precise marginal
98 economic values.

99

100 **Farrow-to-finish (commercial) herd bio-economic model framework**

101 The farrow-to-finish (commercial) herd bio-economic model (Fig. 1 and Supplementary Fig.
102 S1A) manages the complete lifecycle of purchased gilts within a single farm, commencing with
103 gilt acquisition and rearing to breeding age, followed by systematic artificial insemination
104 implementation. Sows progress cyclically through successive reproductive phases—
105 insemination, pregnancy, farrowing, lactation, and mating intervals—until they reach
106 maximum productive longevity, with standardized feeding procedures dynamically adjusted to
107 physiological stage-specific nutritional demands and health status. Throughout this cycle, sows
108 face culling risks due to mortality, disease, no heat, or suboptimal litter performance. Progeny

109 development occurs through three sequential phases: lactation, where piglets derive primary
110 nutrition from suckling while progressively incorporating creep feed; nursery, which follows
111 weaning and facility transfer; and finishing, where pigs advance to target slaughter weight. To
112 reflect operational realities, the nursery and finishing stages may be subdivided into one-to-
113 three management intervals aligned with specific feeding regimens to accommodate evolving
114 nutritional requirements. Slaughter pigs are sold on the basis of live weight, carcass weight, or
115 quality grading according to actual market conditions.

116

117 **Single pure-line breeding system bio-economic model framework**

118 The single pure-line breeding system bio-economic model (Fig. 1 and Supplementary Fig. S1B)
119 achieves within-breed genetic improvement through controlled self-breeding, commencing
120 with gilt acquisition via internal selection or external purchase and progressing through
121 sequential reproductive cycles until maximum productive longevity is reached. The structure
122 of this model is further optimized on the basis of the farrow-to-finish herd model. The difference
123 is that after the nursery period is completed, the pigs are separated into breeding group and
124 fattening group. Within the breeding group, breeders or farmers evaluate pig performance and
125 select the best offspring as replacements for boars and sows, and the superior boars are
126 transferred to AI stations for semen production, while suboptimal candidates join commercial
127 fattening streams. Notably, during the nursery stage and in the breeding group, a certain number
128 of breeding pigs can be sold outside the system for purchase and use by other herds. The
129 unselected gilts/boars are fattened and sold as commercial pigs along with the fattening group,
130 collectively constituting the system's dual-output revenue structure.

131 **Two-way crossbreeding system bio-economic model framework**

132 The two-way crossbreeding system bio-economic model integrates dam breed A and sire breed
133 B within a tiered production structure, synthesizing frameworks from both farrow-to-finish herd
134 and pure-line breeding models (Supplementary Fig. S2A). Nucleus herds maintain purebred
135 populations (A and B), whereas multiplier herds utilize semen transferred from breed B to
136 inseminate breed A dams, generating hybrid progeny (A×B). To account for practical variations
137 in productive lifetime and rearing environments when breed A is utilized for different purposes
138 within breeding programs, individuals designated exclusively for hybrid progeny production
139 were segregated into distinct reproduction A. Within this integrated system, breeds A and B
140 retain their respective reproductive functionalities, with critical directional flows: purebred
141 female offspring of breed A either replenish nucleus herds or transfer to multiplier herds for
142 hybrid production, while male offspring of breed B supply semen for intra-breed artificial
143 insemination or crossbreeding in multiplier herds. Hybrid offspring (A×B) are exclusively used
144 for fattening and sale, which establishes a clear genetic and economic separation between
145 purebred maintenance streams and commercial output streams.

146

147 **Three-way crossbreeding system bio-economic model framework**

148 The three-way crossbreeding system bio-economic model integrates breed A, B and C through
149 a hierarchical genetic pyramid that includes the nucleus, multiplier and commercial herds. This
150 model extends integrated frameworks from farrow-to-finish herd and pure-line system to five
151 distinct populations (Supplementary Fig. S2B): nucleus herds maintaining pure breeds A, B,
152 and C; reproduction A groups producing A×B hybrids using breed A gilts (sourced from the

153 nucleus) and breed B semen; and reproduction AB groups where elite A×B females (selected
154 from reproduction A outputs) are crossed with breed C semen to produce exclusively (A×B)×C
155 finishers. This process establishes a unidirectional genetic flow from nucleus conservation to
156 commercial terminal cross production.

157

158 **Bio-economic model sow herd and offspring structure**

159 On the basis of the framework of each model, the population counts of sows and their offspring
160 at different production stages serve as the fundamental basis for determining total profit.

161 Initially, the sow population is assumed to remain constant and is distributed in dynamic
162 equilibrium across reproductive cycles. In the farrow-to-finish herd model, to minimize the
163 number of model parameters and enhance applicability, sow numbers at subsequent stages are
164 calculated on the basis of stage-specific mortality and culling rates. Using parameters for litter
165 size and stage-specific offspring mortality rates, the model subsequently computes the number
166 of offspring surviving each stage.

167 For other breeding systems, owing to the complexity inherent in breeding systems, the steady-
168 state distribution of sows across various physiological states within different reproductive
169 cycles is calculated using the Markov chain method [32,33]. We adopted the method of
170 constructing the Markov chain transition matrix \mathbf{T} by Wolfová *et al.* [29], which has been
171 successfully applied to calculate the structure of the sow herd. The transition matrix \mathbf{T}
172 represents the sequential transfer process among all parities of the sow before the maximum
173 service life is reached. For each parity, sow states are categorized into mutually exclusive
174 scenarios: sows reaching the maximum number of allowed matings and culled due to

175 conception failure; sows dying during pregnancy or lactation; sows culled during the mating
176 interval for health reasons; and sows advancing to the next breeding cycle. The probabilities of
177 these mutually exclusive scenarios are regarded as elements in the matrix \mathbf{T} . Then, an initial
178 vector \mathbf{I} is established to represent the probability ratio at the time of population establishment,
179 with dimensions equal to the number of rows in matrix \mathbf{T} :

$$180 \quad \mathbf{I} = [1 \ 0 \ 0 \ \dots \ 0]$$

181 Thus, the state distribution \mathbf{P} of the number of sows at time t is expressed as follows:

$$182 \quad \mathbf{P}_t = \mathbf{I}\mathbf{T}^t$$

183 According to the properties of the Markov chain, \mathbf{P} remains stable as time t increases. We
184 set the default value of t to 30 to obtain the probability distribution of sows in the stable state
185 as well as the number of sows in different productive states. The parameters for litter size and
186 stage-specific offspring mortality rates were subsequently used to compute the number of
187 offspring surviving at each stage. These models integrate the growth and reproductive processes
188 of sows and offspring into a unified system, regardless of whether offspring are raised on
189 separate finishing farms.

190

191 **Costs and revenues in bio-economic model**

192 Building upon these population frameworks, the costs and revenues of all the components of
193 the bio-economic models are considered, which are included in the models as follows:

194 Costs include the following:

- 195 • Feed costs for gilts and sows at different stages (e.g., pregnancy, lactation, and the
196 mating interval).

- 197 • Feed costs for piglets during lactation (creep feed).
- 198 • Feed costs for weaned pigs in the nursery and fattening pigs in the feeder-to-finish stage
199 (up to three stages can be considered, depending on actual feeding needs).
- 200 • Feed costs for the breeding group, separated by sex (boars and replacement gilts during
201 the feeder-to-finish stage, with up to three stages considered).
- 202 • Fixed costs for gilts and sows (e.g., medical costs, housing costs, water, electricity,
203 environmental costs, and labor costs).
- 204 • Artificial insemination costs and farrowing costs for gilts and sows.
- 205 • Costs for the disposal of dead animals.
- 206 • Fixed costs for sow offspring at different stages (e.g., testing and evaluation of breeding
207 animals, medical costs, housing costs, water, electricity, environmental costs, and labor costs).
- 208 • Costs for transporting and selling pigs.

209 Revenue includes the following:

- 210 • Revenue from culling sows and selling breeding group pigs.
- 211 • Revenue from selling finished pigs.

212 The data required in the calculation process of these costs and revenues are used as parameters
213 input to the model and must be input by the user.

214

215 **Production efficiency**

216 In farrow-to-finish herds, production efficiency is measured as total profit per finished pig,
217 expressed as follows:

218
$$Prof_f = \frac{Trevenues_f - Tcosts_f}{\text{number of finished pigs}}$$

219 where $Prof_f$ is the total profit per finished pig and where $Trevenues_f$ is the total revenue,
220 including revenue from culling sows and finished pigs for sale. $Tcosts_f$ is the total cost, which
221 includes all feed costs and non-feed costs for the sows and their offspring.

222 In other breeding systems, production efficiency is expressed as the total profit per sow per
223 parity as follows:

224
$$Prof_s = \frac{Trevenues_s - Tcosts_s}{\text{number of sows}}$$

225 where $Prof_s$ is the profit per sow in this system and where $Trevenues_s$ is the total revenue,
226 including revenue from culling sows, breeding group pigs and finished pigs for sale. $Tcosts_s$
227 is the total cost, which includes all feed costs and non-feed costs for the sows and their offspring.

228

229 **Calculation of NDEs by using the gene flow method**

230 Conceptually, the gene flow method is designed to mirror the biological lifecycle of the swine
231 herd by tracking gene transmission through two fundamental processes: aging (growth) and
232 reproduction. To operationalize this, the population was stratified into distinct sex-age groups
233 with a discrete time step of half a year, corresponding to the average reproductive cycle of sows.

234 According, a gene transmission matrix \mathbf{P} was constructed based on the breeding system's
235 structure, where rows represent sex-age groups at time t and columns represent groups at $t -$
236 1.

237 For the pure-line bio-economic model (Fig. 2A), $\mathbf{P1}$ is partitioned into four quadrants to
238 capture specific transmission pathways: the upper-left and lower-right quadrants represent
239 within-sex transmission (sire-to-sire and dam-to-dam), while the off-diagonal quadrants

240 represent cross-sex transmission (sire-to-dam and dam-to-sire). Each quadrant integrates two
241 matrices: reproduction (**P2**) and ageing (**P3**). Within the reproduction matrix **P2**, the elements
242 in the first row (represented by asterisks, *) quantify the proportion of genes transmitted from
243 parental group 1-x at $t - 1$ to the offspring (age group 1) at time t ; these values range from
244 0 to 0.5, representing the Mendelian inheritance probability scaled by the reproductive
245 contribution of the respective age group. Conversely, the growth matrix **P3** contains identity
246 elements ("1") on the sub-diagonal, representing the survival and transition of animals to the
247 subsequent age class. This framework extends to crossbreeding systems by integrating multiple
248 pure-line dynamics. For the two-way crossbreeding system (Fig. 2B), the matrix tracks gene
249 flow through maternal line A and paternal line B, explicitly modeling the production of hybrid
250 offspring. Owing to age-dependent utilization differences in maternal line A, the reproduction
251 A gene transmission process is modeled separately. The complete gene flow matrix integrates
252 submatrices **P1**, **P2**, **P3**, and **P4**, with all remaining non-functional elements set to 0.
253 Similarly, in the three-way crossbreeding system (Fig. 2C), reproduction AB is modeled
254 independently to account for age-specific usage, with the global transmission matrix
255 constructed to strictly align with the artificial insemination protocols and the hierarchical flow
256 of the crossbreeding program.

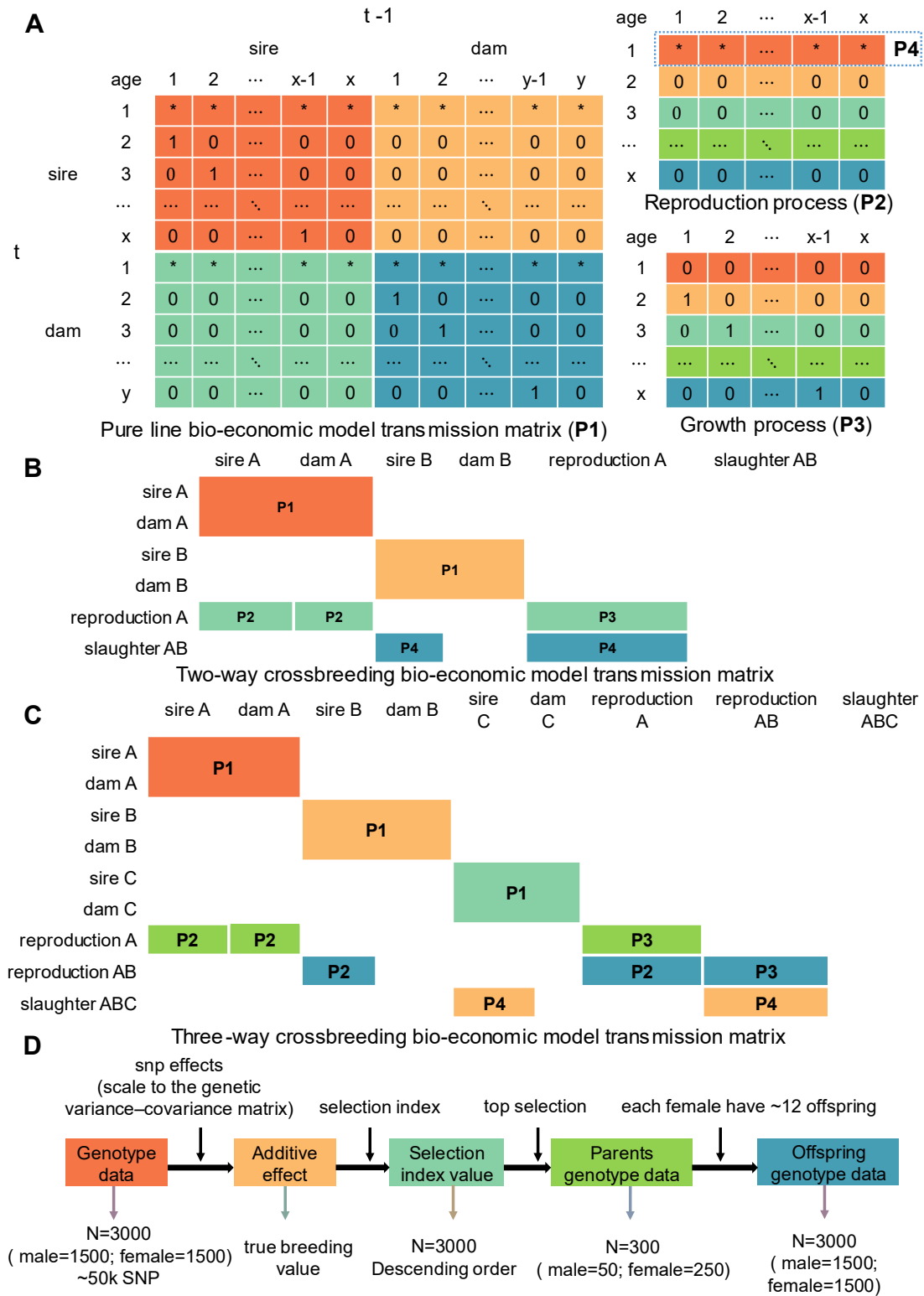
257 To calculate the gene expression of the selected breeding pigs, a gene probability vector **m**
258 (dimensions matching the number of age–sex groups) was defined. At $t = 0$, the element
259 corresponding to the initial age–sex group of the selected pigs was assigned a value of 1, with
260 all other elements initialized to 0. The temporal probability distribution of genes across age–
261 sex groups was derived iteratively using the following equation:

262
$$\mathbf{m}_t = \mathbf{P}\mathbf{m}_{t-1}$$

263 where \mathbf{m}_t represents the gene probability vector at time t . Traits expressed earlier in the
264 breeding period carry greater economic weight than those expressed later. A discount factor r
265 was therefore applied to account for the temporal value of breeding investments. For each trait,
266 a realization vector \mathbf{h} (dimensions matching the number of age–sex groups) was defined to
267 represent trait expression levels across sex–age groups. Its elements denote the proportion of
268 pigs expressing this trait in each age–sex group of pigs relative to the total number of sows.
269 This vector \mathbf{h} was assumed to be constant throughout the investment period. The NDEs for a
270 trait are calculated as follows:

271
$$n_{ij} = \sum_{t=1}^D \mathbf{h}'_i \mathbf{m}_{jt} (1+r)^{-t}$$

272 where n_{ij} represents the NDEs of trait i in sex group j ; D represents the breeding investment
273 period; \mathbf{h}_i represents the realization vector for trait i ; \mathbf{m}_{jt} represents the gene probability
274 vector for sex group j at time t ; r represents the discount factor; and t represents the time
275 of trait expression.



276

277 **Figure 2.** Structure of gene transmission matrices and the simulation flowchart. (A) The

278 transmission matrix (P1) for the pure-line bio-economic model. The matrix models the gene

279 flow between sex-age groups over a time step of half a year (representing one age class).

280 Columns represent the population state at time $t - 1$, and rows represent time t . The matrix
281 is divided into four quadrants representing genetic transmission pathways (sire to sire, sire to
282 dam, dam to sire, dam to dam). The first row of each quadrant (marked with asterisks, *)
283 represents the genetic contribution of parents to offspring (values 0–0.5, reflecting Mendelian
284 inheritance). The sub-diagonal elements marked with "1" represent the aging process, where
285 surviving individuals move to the next age class. (B) The transmission matrix for the two-way
286 crossbreeding system. This matrix integrates the pure-line dynamics for Dam A and Sire B,
287 modeling the flow into Reproduction A and the terminal Slaughter AB group. (C) The
288 transmission matrix for the three-way crossbreeding system. This extends the model to include
289 Sire C, producing the final Slaughter ABC population through sequential crossbreeding steps.
290 (D) Flowchart of the multi-generational simulation. The process illustrates the calculation steps
291 from initial genotype data to the calculation of additive effects, selection index values, and the
292 selection of top parents to generate the next generation's offspring genotype data.

293

294 **Economic weights and sensitivity analysis**

295 The MEV of a trait is calculated as the change in profit per unit change in the trait mean (TM),
296 corresponding to the partial derivative of the profit function. In this study, this value is
297 approximated via a central finite difference method:

$$298 \quad MEV = \frac{\partial Prof}{\partial TM} \approx \frac{Prof_{(1+0.1\%)TM} - Prof_{(1-0.1\%)TM}}{0.2\%TM}$$

299 where $Prof_{(1+0.1\%)TM}$ and $Prof_{(1-0.1\%)TM}$ represent profits when the trait mean is
300 increased or decreased by 0.1%, respectively. A variation of $\pm 0.1\%$ was chosen to minimize
301 discretization error, ensuring the result accurately approximates the tangent slope of the profit

302 function at the population mean. In farrow-to-finish herds, the MEV serves as a direct proxy
303 for trait weight in the selection index. For other breeding systems, the economic weight (EW_i)
304 of trait i is derived by scaling its MEV by the NDE (n_{ij}) associated with the selected sex group,
305 following $EW_i = MEV_i \times n_{ij}$.

306 To explore the sensitivity of economic weights to such fluctuations, three simulation scenarios
307 (feed price fluctuations, non-feed price fluctuations, and sale price fluctuations) were set up to
308 predict the effects of different types of price fluctuations on economic weights. In each
309 simulated scenario, the economic weights of target traits under incremental price increases of
310 10%, 20%, 30%, 40%, and 50% can be iteratively calculated via a bio-economic model.

311

312 **Relative economic weights and standardized selection index**

313 To compare the relative importance of different traits, standardized economic weights (SEWs)
314 were first calculated by multiplying each trait's economic weight (EW) by its genetic standard
315 deviation. The relative economic weight (REW) was expressed as the percentage contribution
316 of a trait's SEW to the total SEWs of all the traits:

$$317 \quad SEW_i = EW_i \times \sigma_i$$

$$318 \quad REW_i = \frac{SEW_i}{\sum_1^n SEW_i} \times 100\%$$

319 where σ_i is the genetic standard deviation of trait i and where n is the total number of traits.

320 In the context of modern breeding programs, the selection index (I) is first calculated as the
321 linear combination of EW and EBV :

$$322 \quad I = \sum_{i=1}^n (EW_i \times EBV_i)$$

323 Unlike the classical index which relies on phenotypic to infer genetic merit, this approach
324 utilizes pre-estimated EBVs directly. Since EBVs are now directly computable via upstream
325 evaluations (e.g., Pedigree-BLUP or GBLUP), the construction of a phenotype-based index
326 (and its associated covariance estimation) is rendered unnecessary. Subsequently, to facilitate
327 practical decision-making, the mean and variance of this selection index are standardized (H)
328 and expressed as follows:

$$329 \quad H = \mu + \frac{I}{\sqrt{EW'(V)EW}} \times STD$$

330 where μ is the average of the standardized selection index (e.g., 100), V is the variance–
331 covariance matrix of the EBVs, and STD is the standard deviation of the standardized
332 selection index (e.g., 25).

333

334 **Simulation of multi-generational selection response**

335 The selection response module is validated using a built-in simulated pig population containing
336 3,000 genotyped individuals (~50K SNP density) with full pedigree records. This scale
337 effectively represents a nucleus breeding herd and allows for rapid simulation of genetic gain.
338 This synthetic population, adapted from our previous work [34], replicates realistic genetic
339 architectures, trait correlations, and population structures observed in real pig herds. To predict
340 multi-generation genetic gain, the simulation workflow proceeded as follows (Fig. 2D): it was
341 assumed that target traits were affected by a certain number of QTLs per chromosome, and
342 additive genetic effects for QTLs were sampled from a multivariate normal distribution scaled
343 to match the user-defined genetic variance–covariance matrix. For each generation, candidate
344 pigs were ranked by the calculated selection index, derived from true breeding values (TBVs)

345 to validate the theoretical efficiency of the index. The top 50 males and 250 females were
346 selected as parents for the next generation, and each female produced 12 offspring (sex ratio of
347 0.5), maintaining a population size of 3,000 per generation. For each generation, pedigrees,
348 genotypic additive effects, and indices were recorded, and the means of additive effects and
349 selection indices were visualized as line graphs to track the multi-generation selection response
350 throughout the future breeding cycles/generations. This simulation is explicitly designed to
351 allow users to rapidly validate the expected genetic gain of the index on standard personal
352 computers, balancing statistical representativeness with operational efficiency.

353

354 **QTL allelic effect values**

355 The built-in genotype data encompass 18 chromosomes, with each chromosome associated with
356 40 quantitative trait locus (QTL) positions. First, the genotype data are read to extract the
357 genomic positions of both the QTLs and the markers. The effect value for each QTL locus is
358 subsequently simulated using a multivariate normal distribution. Let the vector \mathbf{X} represent
359 the effect of each QTL on n traits, following a multivariate normal distribution $\mathbf{X} \sim N_n(\boldsymbol{\mu}, \mathbf{V})$,
360 where $\boldsymbol{\mu}$ represents the mean vector and \mathbf{V} represents the genetic variance–covariance matrix
361 of n traits. Then, N ($QTL \times chromosome\ number$) independent and identically distributed
362 random samples are generated, and the locus effect values matrix \mathbf{A} is obtained:

363

$$\mathbf{A} = \begin{bmatrix} \mathbf{X}_1^T \\ \mathbf{X}_2^T \\ \vdots \\ \mathbf{X}_N^T \end{bmatrix}_{N \times n}$$

364 where $\mathbf{X}_i \sim N_n(0, \mathbf{V})$ and $i = 1, 2, \dots, N$. Finally, the TBVs for the traits in the base population
365 are computed by multiplying the genotype dosage matrix (sum of alleles 1 and 2 at each QTL)

366 by the QTL effect values matrix, with these effects subsequently standardized to align with the
367 target population's genetic variance–covariance matrix. Scaling is performed as follows:

$$368 \quad \text{Scale} = \frac{\text{sqrt}(\text{Input variance matrix})}{\text{sqrt}(\text{TBVs variance matrix})}$$

369 then, the QTL allelic effect at each locus is scaled as $(\text{allelic effect} - \text{QTL mean}) \times$
370 *Scale*. The effect values after scaling are used to calculate the subsequent additive genetic
371 effect.

372

373 **Software architecture and system requirements**

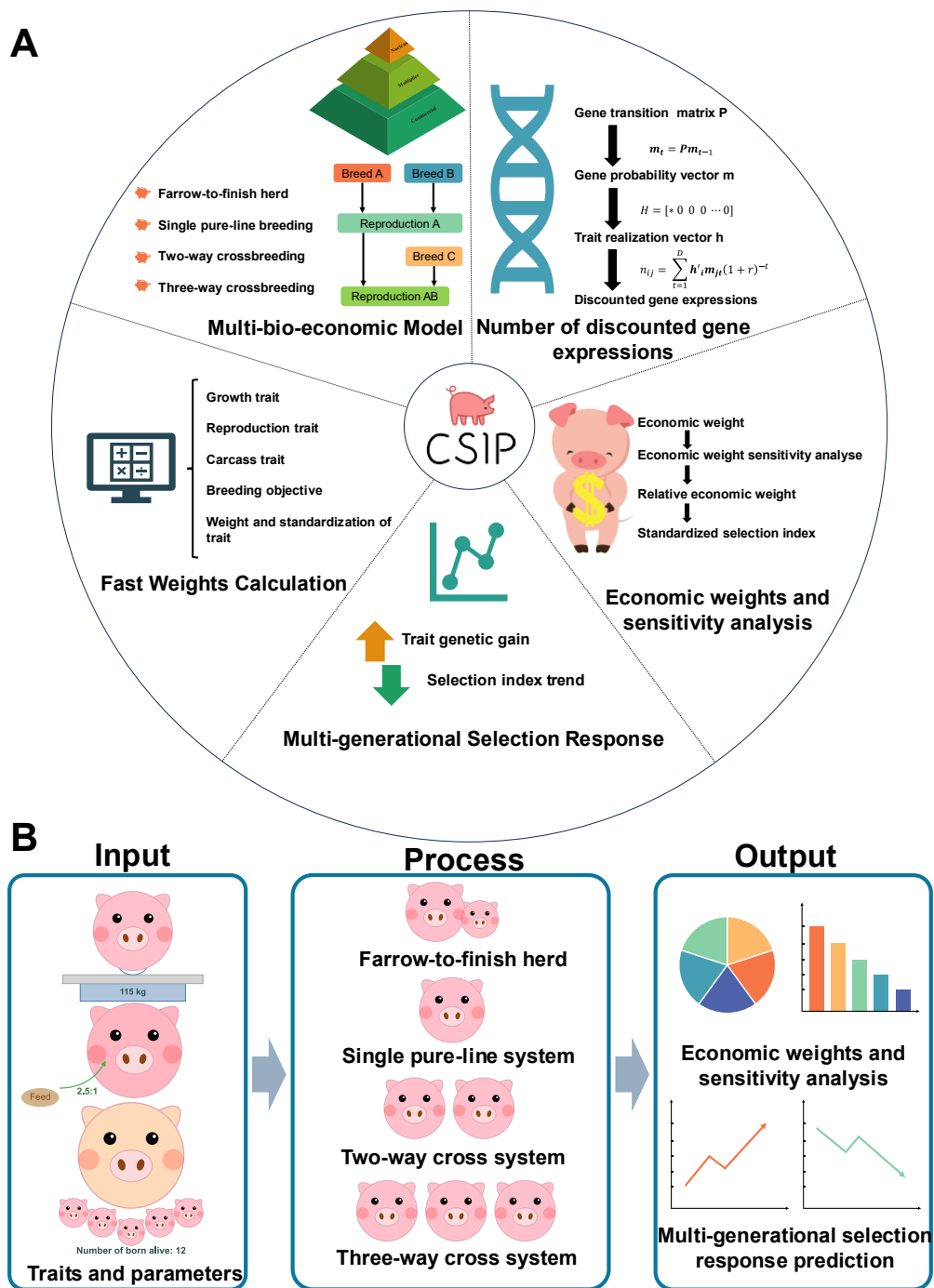
374 The software was developed using a multi-tier architecture. The frontend was implemented
375 with *Vue.js*, which is a responsive and intuitive interface that facilitates seamless interaction
376 with various modules and visualizations. The backend computational engine was developed in
377 *Python*, with *NumPy* and *Pandas* used for efficient numerical processing and data manipulation
378 and *SciPy* used for advanced scientific computations. *Electron* was used to package the
379 application, and excellent cross-platform capabilities were ensured.

380

381 **Selection index for pig production in the farrow-to-finish (commercial) herd and three-** 382 **way crossbreeding system**

383 We use the farrow-to-finish (commercial) herd model to demonstrate the functions of the CSIP
384 software. A dam selection index with five traits (age at target weight (ATW), finishing-phase
385 feed conversion ratio (F-FCR), backfat depth (BFD), loin muscle depth (LMD), and number of
386 born alive (NBA)) was formulated, and the multi-generational genetic response was simulated.
387 Moreover, a three-way crossbreeding system model, incorporating genetic data from three

388 nucleus populations (Yorkshire, Landrace, and Duroc) and economic parameters derived from
389 typical commercial production systems and market surveys, was employed to calculate
390 economic weights and design indices to demonstrate that the software can calculate the
391 economic weights of up to 26 traits and the selection index of the crossbreeding system. The
392 input parameters that generate these results can be found in Supplementary Table S1–6.



393

394 **Figure 3.** Overview of the CSIP software. (A) Core functional modules. The diagram illustrates

395 the five interconnected computational components integrated within CSIP: the multi-bio-

396 economic model, number of discounted gene expressions, economic weights and sensitivity

397 analysis, multi-generational selection response, and fast weights calculation. (B) Operational

398 workflow. The diagram presents the data flow from the input of traits and parameters, through
399 the Process of breeding system, to the output of visualized analysis results.

400

401 **Results**

402 **Overview of the CSIP software**

403 CSIP is specialized software for formulating pig selection index via the integration of bio-
404 economic models and multi-generational prediction of selection response within a streamlined,
405 practical, and fully visualized workflow. Its core comprises five interconnected modules (Fig.
406 3A): the multi-bio-economic model, number of discounted gene expressions, economic weights
407 and sensitivity analysis, multi-generational selection response, and fast weights calculation.

408 The multi-bio-economic model module provides selectable frameworks (e.g., farrow-to-finish
409 (commercial) herd, single pure-line, two-way cross, and three-way cross breeding systems) for
410 calculating the MEVs of traits. Each model integrates 26 economic productive traits, including
411 growth, reproduction, and carcass traits. The number of discounted gene expressions module
412 uses a gene flow approach to compute NDEs. The obtained MEVs and NDEs are then integrated
413 into the economic weights and sensitivity analysis module to derive economic weights and
414 analyze the sensitivity of these weights to fluctuations in economic parameters. This module
415 also supports the calculation of relative economic weights alongside the formulation of a
416 standardized selection index. The multi-generational selection response module allows the
417 prediction of multi-generational selection response when the designed selection index is applied
418 in the breeding system. This approach allows a comparison of the consequences of genetic gain
419 when a suboptimal selection index is applied in a breeding project. The fast weights calculation

420 module provides alternative flexible index calculation methods. In this approach, economic
421 weights are derived on the basis of the desired genetic gains or simplified economic values
422 calculated by using few input parameters. These modules provide a comprehensive solution for
423 pig selection index formulation.

424 To ensure an enhanced user experience, the software is built using Electron, Vue.js and Python,
425 enabling excellent cross-platform capabilities. CSIP streamlines the operation into a
426 straightforward 'input-process-output' process (Fig. 3B). Each analytical module is guided
427 through a step-by-step workflow, effectively eliminating the need for traditional command line
428 interfaces and complex file parsing procedures. Demonstrating high computational efficiency
429 on standard personal computers (Intel Core Ultra 7 265K processor, 32 GB RAM), CSIP
430 completes economic weights calculation in seconds and executes stochastic validation
431 (simulating a typical 3,000-head nucleus herd over 10 generations) in approximately 8 minutes.

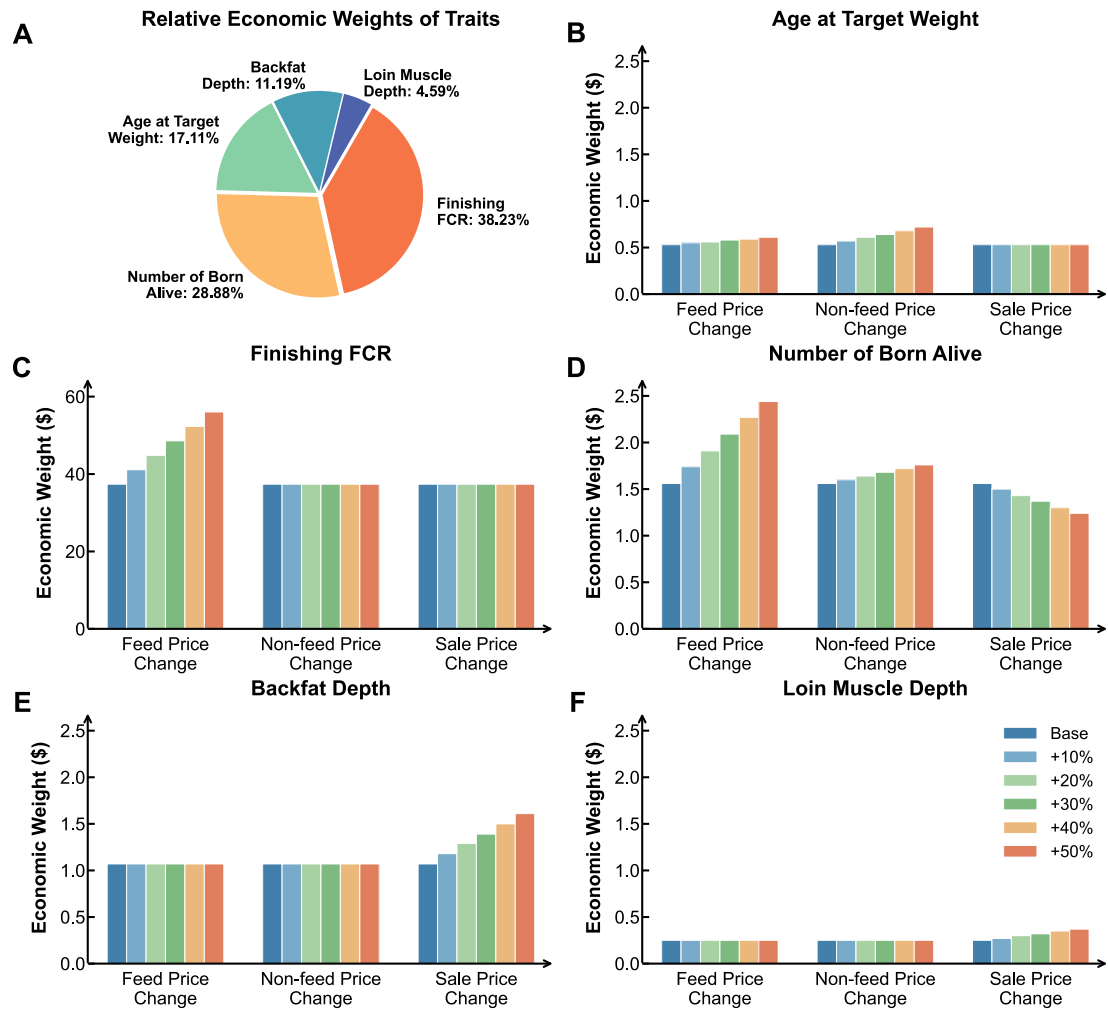
432 Methodologically, the software covers diverse breeding structures and enhances the precision
433 of economic weight calculations by synergistically integrating a gene flow approach for
434 calculating NDEs directly within specialized breeding system bio-economic models and
435 facilitates the computation of economic weights for sensitivity analysis, relative economic
436 weights and standardized selection index. Furthermore, CSIP features an innovative module for
437 the prediction of multi-generational selection response and is proactively designed to mitigate
438 antagonistic trait response and suboptimal genetic gain arising from poorly designed breeding
439 programs. Overall, this integrated platform represents the first full-process visualized software
440 for the design and formulation of the pig selection index.

441 **Results of the economic weight, selection index and sensitivity analysis for the farrow-to-**
442 **finish herd**

443 The relative economic weights are shown in Fig. 4A, where F-FCR was prioritized as the most
444 critical trait (38.23%), followed by NBA (28.88%), ATW (17.11%), BFD (11.19%), and LMD
445 (4.59%). In the current baseline scenario (designed selection index), economic weights were
446 calculated to be \$0.53 per day for ATW, \$37.37 per unit for F-FCR, \$1.56 per piglet for NBA,
447 \$1.07 per millimeter for BFD, and \$0.25 per millimeter for LMD (Supplementary Table S7).
448 For the sensitivity analyses, when feed prices increased within a certain range, economic
449 weights increased for ATW, F-FCR, and NBA (Fig. 4B-D); when the non-feed price increased,
450 only ATW and NBA presented higher economic weights, whereas other traits remained
451 unchanged (Fig. 4B, D); when the sale price increased, higher sale prices increased the
452 importance of BFD and LMD (Fig. 4E, F) while decreasing the weight of NBA (Fig. 4D). These
453 trends demonstrate the sensitivity of economic weights to market fluctuations, providing
454 actionable insights for dynamic breeding program adjustments. The standardized selection
455 index was scaled to a mean of 100 and a standard deviation of 25 and is expressed as follows:

$$456 \quad H = 100 - 101.75 \times EBV_{F-FCR} - 1.44 \times EBV_{ATW} + 8.50 \times EBV_{NBA} - 2.91 \times EBV_{BFD}$$
$$457 \quad \quad \quad + 0.68 \times EBV_{LMD}$$

458 where *EBV* represents the estimated breeding value for the corresponding trait. This
459 standardized selection index can be directly applied in actual production, significantly lowering
460 the threshold for formulating the selection index.



461

462 **Figure 4.** Relative economic importance and sensitivity analysis of traits in the farrow-to-finish

463 system. (A) Pie chart showing the relative economic weights of the five evaluated traits. (B–F)

464 Sensitivity analysis of economic weights for (B) Age at Target Weight, (C) Finishing FCR, (D)

465 Number of Born Alive, (E) Backfat Depth, and (F) Loin Muscle Depth. The "Base" scenario

466 represents the reference economic weights derived for the originally designed selection index

467 (under current market parameters), while the gradient bars illustrate fluctuations resulting from

468 stepwise increases (+10% to +50%) in feed prices, non-feed prices, and sale prices.

469

470 **Results of multi-generational selection response under the designed selection index and**
471 **price variation scenarios for the farrow-to-finish herd**

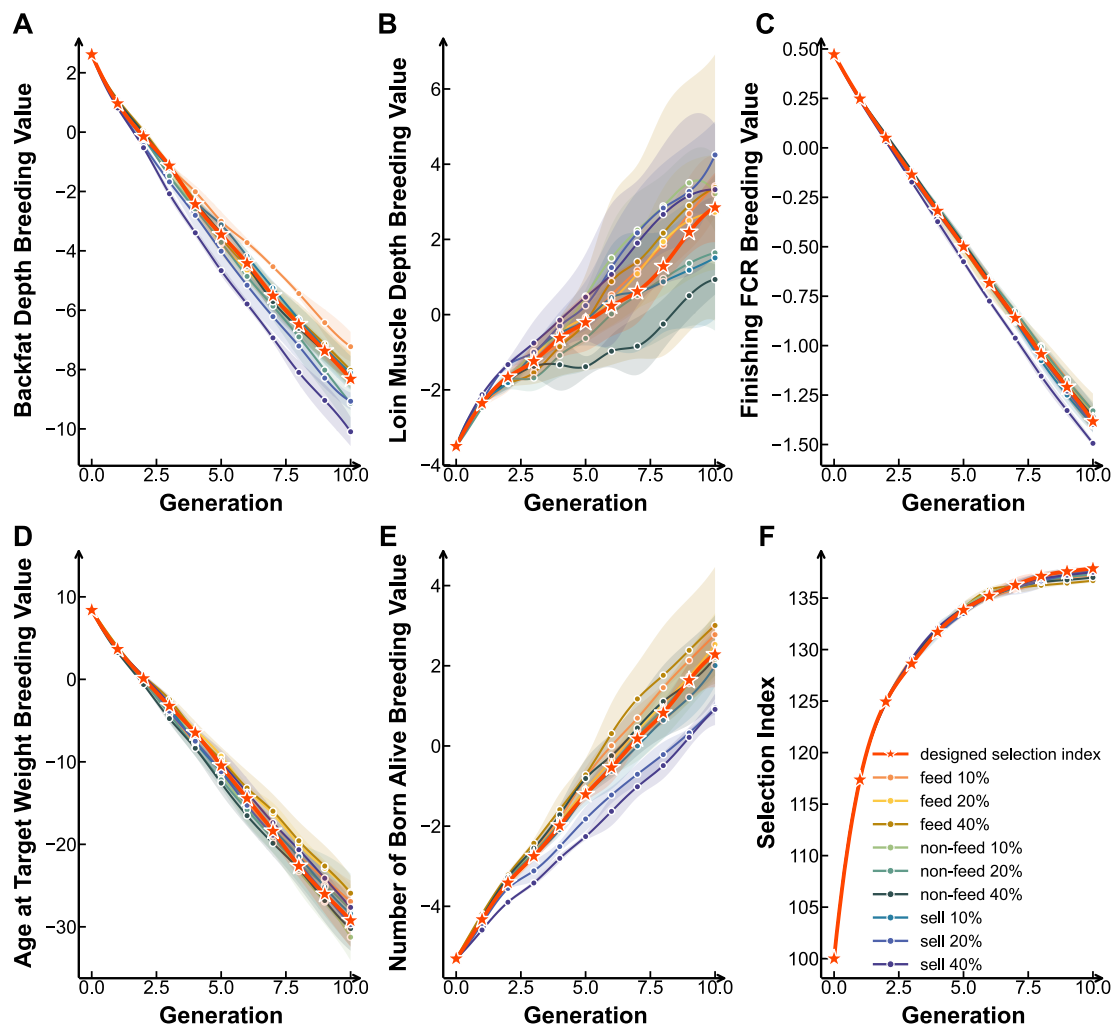
472 To assess the impact of economic fluctuations on economic weights and selection response,
473 relative economic weights were calculated in the designed selection index scenario and
474 scenarios with the price increases of 10%, 20% and 40% (Supplementary Table S8), and the
475 corresponding genetic gains were subsequently computed (Supplementary Fig. S3-5). The
476 results revealed that ATW, F-FCR and BFD tended to decrease, whereas NBA and LMD tended
477 to increase (Fig. 5A–E). Market fluctuations had a limited effect on the selection response, as
478 the trends of different lines are similar with different price variations. Although the genetic
479 gains of certain traits increased slightly in the fluctuating price scenarios, others showed
480 marginal decreases compared with the designed selection index. The overall trend of the
481 selection index values remained largely consistent across scenarios with rapid increases in the
482 early generations (generations 1–3), followed by a gradual plateau in generations 7–10, but the
483 design selection index was the highest selection index in the later stage (Fig. 5F), indicating the
484 robustness and stability of the formulated selection index in the face of moderate economic
485 variability.

486

487 **Results of the tenth-generation multi-generational selection response obtained with**
488 **different selection strategies in the farrow-to-finish herd**

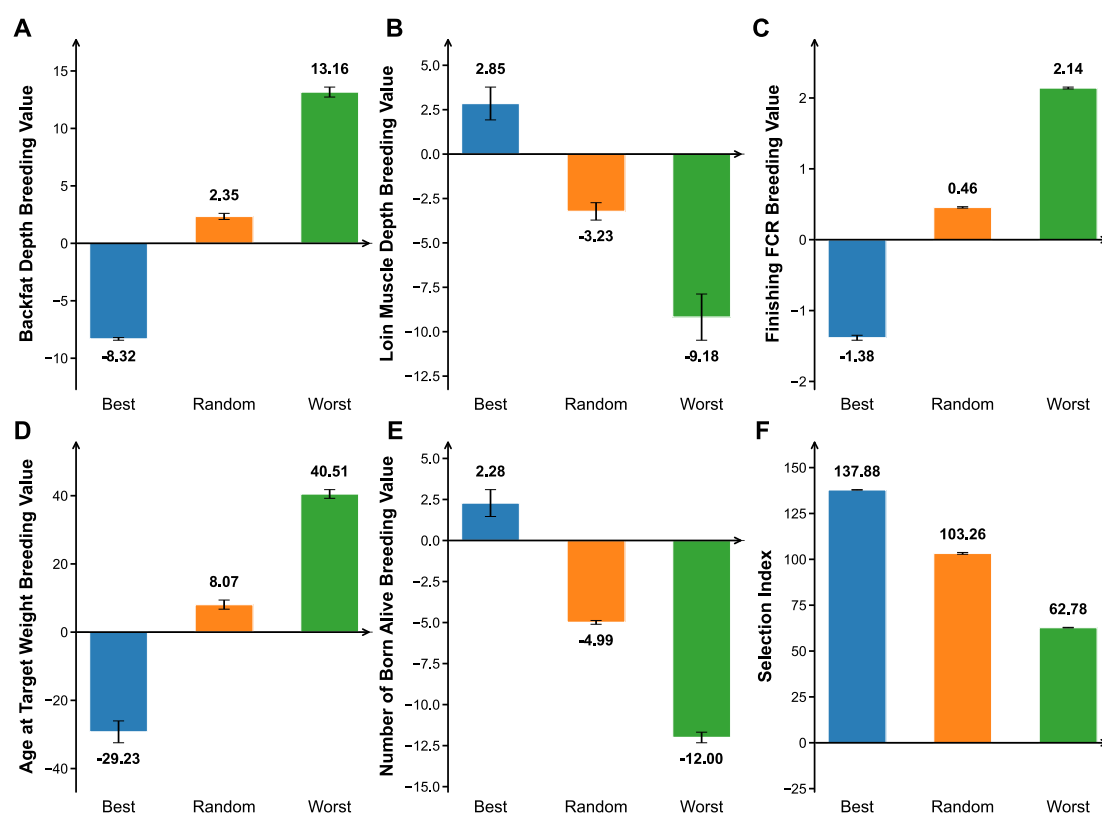
489 To evaluate the effectiveness of the formulated selection index, multi-generational selection
490 response simulations were performed under three selection strategies: best selection, random
491 selection, and a worst-case selection scenario. The results from the 10th generation (Fig. 6)

492 demonstrated that the index formulated by the CSIP software (the best selection scenario)
493 significantly improved the genetic value of the target traits and achieved high overall genetic
494 gains in the desired direction. The worst selection strategy led to a pronounced decline in
495 performance for all the traits. These results highlight the effectiveness of the software-
496 formulated selection index in achieving targeted genetic improvement, underscoring its
497 potential to increase both genetic gain and overall performance in pig breeding programs.
498



499
500 **Figure 5.** Predicted multi-generational genetic gain and selection index trends under varying
501 economic scenarios. (A–E) Cumulative genetic gain for traits. The trajectories display the True

502 Breeding Values (TBVs) over 10 generations for (A) Backfat Depth, (B) Loin Muscle Depth,
503 (C) Finishing FCR, (D) Age at Target Weight, and (E) Number of Born Alive. Note that the
504 occurrence of negative TBVs is inherent to the simulation design, as multi-trait SNP effects
505 were initially sampled from a distribution with a mean of zero. (F) Trend of the Selection Index.
506 This panel illustrates the overall response of the aggregate breeding objective. Scenarios: The
507 orange line with star markers represents the baseline response using the designed selection
508 index (under current economic parameters). The other colored lines represent sensitivity
509 scenarios with varying price increments. Shaded regions indicate the standard deviation of the
510 simulation.



511
512 **Figure 6.** Comparative analysis of tenth-generation genetic response and selection index values
513 under contrasting selection strategies. (A–E) Cumulative True Breeding Values (TBVs) for
514 target traits. The bar charts display the final genetic status at generation 10 for (A) Backfat

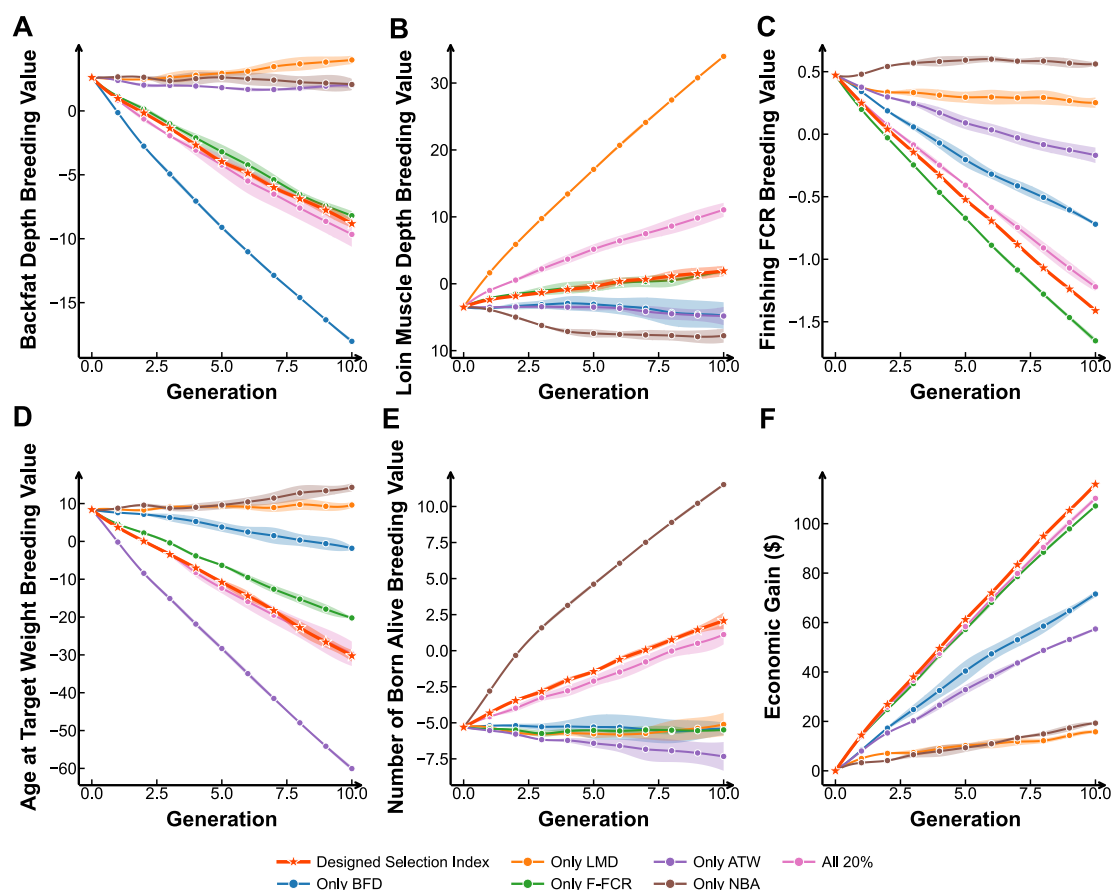
515 Depth, (B) Loin Muscle Depth, (C) Finishing FCR, (D) Age at Target Weight, and (E) Number
516 of Born Alive. (F) Selection Index. Strategies: The analysis contrasts three selection strategies:
517 Best (selection of top-ranking candidates), Random (random mating), and Worst (selection of
518 lowest-ranking candidates). Note: Consistent with the simulation design (initial SNP effects
519 sampled with a mean of zero), negative TBVs for Backfat Depth, Finishing FCR, and Age at
520 Target Weight in the "Best" group indicate a favorable genetic improvement. Conversely,
521 positive values for Loin Muscle Depth and Number of Born Alive represent favorable gains.
522 Error bars indicate the standard deviation of the simulation.

523

524 **Results of the multi-generational selection response and economic gain obtained with**
525 **different economic weights in the farrow-to-finish herd**

526 To evaluate the additional economic benefits of the designed selection index, simulations were
527 conducted in several comparative scenarios, including a designed selection index, selection
528 based on a single trait alone, and uniform relative economic weights assigned to all the traits.
529 The results demonstrated that the targeted trait presented a higher genetic gain in single-trait
530 selection than in the other scenarios (Fig. 7A–E); however, the genetic trends of the nontarget
531 traits either slightly changed, fluctuated, or even were negative, indicating undesirable
532 correlated responses. In contrast, the other scenarios achieved genetic gains in the expected
533 direction across all the traits, although the magnitude of response varied due to differences in
534 economic weights (Fig. 7A–E). In terms of additional economic gain per animal, the single-
535 trait selection scenarios resulted in lower profits than the other strategies did (Fig. 7F). Notably,
536 the designed selection index produced the highest cumulative profit in the later generations

537 across all the evaluated strategies (Fig. 7F), underscoring its effectiveness in achieving balanced
538 genetic improvement and maximizing multi-generational economic return in pig breeding
539 programs.



540
541 **Figure 7.** Comparative evaluation of multi-generational genetic response and economic gain
542 under distinct selection strategies. (A–E) Trajectories of True Breeding Values (TBVs). The
543 plots illustrate the genetic trends over 10 generations for (A) Backfat Depth, (B) Loin Muscle
544 Depth, (C) Finishing FCR, (D) Age at Target Weight, and (E) Number of Born Alive. Note:
545 Consistent with the simulation design, negative trends for Backfat Depth, Finishing FCR, and
546 Age at Target Weight indicate favorable genetic improvement. (F) Economic Gain. This panel
547 quantifies the additional monetary benefit per animal, derived from the change in the additive
548 breeding values of the five target traits (assuming other non-target traits remain constant).

549 Scenarios: The "Designed Selection Index" (orange line with star markers) represents the
550 breeding objective set by the software. "Only [Trait]" (e.g., "Only BFD") refers to single-trait
551 selection strategies. The "All 20%" scenario represents a balanced index strategy where equal
552 relative economic weights (20%) are assigned to each of the five target traits. Shaded regions
553 indicate the standard deviation of the simulation.

554

555 **Results of the economic weights, relative economic weights and multi-generational**

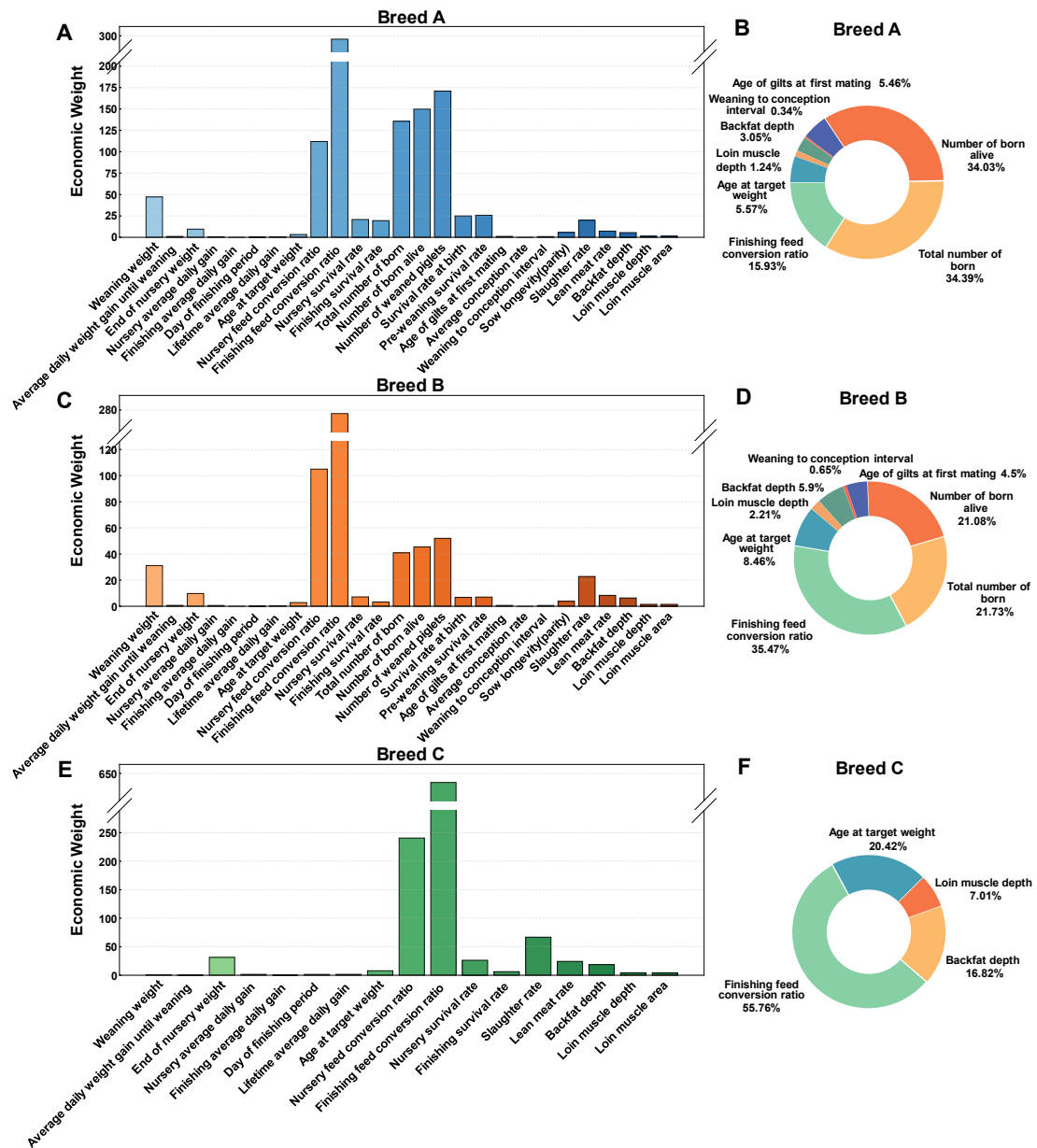
556 **selection response in the three-way crossbreeding system**

557 The economic weights of all traits supported by the software were calculated using a three-way
558 crossbreeding model for three different breeds. The results (Fig. 8A, C, E) indicate that there
559 are notable differences in the economic weights of the same traits between breeds. These
560 variations are attributed to differences in population size and the roles of the breeds within the
561 crossbreeding system. This finding also suggests that considering only the economic values of
562 traits within the commercial herd, without accounting for the structure of the breeding system,
563 may lead to certain inefficiencies. Four production traits and four reproductive traits were
564 subsequently selected from these economic weights to compute the relative economic weights
565 and multi-generational selection response for the three breeds. Breed C, serving as the terminal
566 sire, was considered for production traits only. The results show that relative economic weights
567 differ across breeds within the same system (Fig. 8B, D, F). Furthermore, improper trait
568 selection may result in a slow selection response for some traits and even lead to selection
569 response in the opposite direction of the desired outcome (Supplementary Fig. S6). Therefore,

570 predicting multi-generational selection response while formulating a selection index is crucial

571 to ensure effective genetic improvement.

572



573

574 **Figure 8.** Economic weights and relative economic importance of breeding objective traits for

575 the three-way crossbreeding system. The bar charts display the calculated economic weights

576 for traits in (A) Breed A, (C) Breed B, and (E) Breed C. The pie charts illustrate the proportional

577 contribution of each trait to the overall breeding objective for (B) Breed A, (D) Breed B, and
578 (F) Breed C.

579

580 **Discussion**

581 In this study, CSIP, software for calculating the pig selection index and predicting multi-
582 generational selection response, was developed. The software integrates multiple bio-economic
583 models to address the requirements for formulating a pig selection index and innovatively
584 integrates a simulated selection process to predict multi-generational selection response,
585 enabling users to adjust trait weights on the basis of predicted genetic trends. Furthermore, CSIP
586 provides an intuitive workflow-driven interface lowers the learning barrier, facilitates rapid
587 adoption by interdisciplinary users, and advances data-driven selection index design in swine
588 breeding.

589 In developing bio-economic models for diverse swine breeding systems, our approach
590 incorporates artificial insemination technology as a foundational element. This reflects standard
591 practices in global breeding operations where large-scale breeding and disease control are
592 paramount. To model the growth process of pigs, we utilize fundamental feed intake and price
593 parameters to calculate production costs rather than implementing energy/protein deposition
594 models. While some studies [31,35-37] have integrated detailed growth models into bio-
595 economic frameworks—enabling precise nutritional requirement calculations—such
596 approaches necessitate the development of specific growth models tailored to particular breeds
597 and production conditions. This requirement can be challenging for the majority of farms,

598 thereby constraining the model's applicability. Consequently, our simplified method enhances
599 practical adoption while maintaining economic relevance.

600 The multi-generational selection response quantifies the expected genetic gain of a breeding
601 program and serves as a critical metric for evaluating its design efficacy [38-40]. To model this
602 response, we implemented a simulation framework that replicates multi-generational selection.
603 This framework calculates TBVs on the basis of population genetic parameters, computes
604 selection index values for all candidates, ranks individuals by these values, and selects the top
605 performers. The process iterates over 10 simulated generations, projecting cumulative genetic
606 change. This simulation-based approach—notably absent in conventional selection index
607 methods—addresses a crucial practical gap in breeding program design. Integrating this
608 predictive capability during index development increases formulation robustness by mitigating
609 suboptimal genetic gain arising from antagonistic trait relationships [30].

610 Consequently, this framework offers a validated bio-economic basis for broader breeding
611 scheme design. While established deterministic software (e.g., ZPLAN, SelAction) is widely
612 utilized for optimizing population structures, these tools require reliable economic parameters
613 as inputs. By internally calculating and verifying these weights through the simulation of
614 cumulative genetic gain, our system ensures that the inputs for such deterministic optimizations
615 are biologically sound. This enables a synergistic workflow where the breeding objective is first
616 established, followed by structural refinement using deterministic tools.

617 In future developments, CSIP aims to transition towards a modular, plugin-based architecture,
618 enabling the integration of user-defined algorithms and customizable components [41].

619 Concurrently, the software will be optimized to expand trait coverage and implement AI-driven

620 weight recommendations, particularly for non-economic traits. By combining architectural
621 flexibility with intelligent modeling, CSIP bridges the gap between theoretical frameworks and
622 practical application, thereby advancing precision livestock breeding.

623

624 **Data availability**

625 The software of CSIP can be research freely used at <https://txiang-lab.github.io/CSIP>.

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632 **Conflicts of interest**

633 The authors declare no competing interests.

634

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