

Supplementary Information for

Structural basis for electromechanical coupling and pore block

of human Ca_v1.4

Yitong Sun^{1,4}, Tong Wu^{1,4}, Tongtong Wang¹, Huan Wang¹, Jian Huang²,

Nieng Yan^{1,2,3,5} and Zhangqiang Li^{1,5}

¹State Key Laboratory of Membrane Biology, Beijing Advanced Innovation Center for Structural Biology, Tsinghua-Peking Joint Center for Life Sciences, School of Life Sciences, Tsinghua University, Beijing 100084, China

²Institute of Bio-Architecture and Bio-Interactions (IBABI), Shenzhen Medical Academy of Research and Translation, Guangming District, Shenzhen 518107, Guangdong Province, China

³Institute of Chemical Biology (ICB), Shenzhen Bay Laboratory, Guangming District, Shenzhen 518132, Guangdong Province, China

⁴These authors contributed equally to this work

⁵To whom correspondence should be addressed: N. Yan (nyan@tsinghua.edu.cn) or Z. Li (lizhangq@tsinghua.edu.cn).

This PDF file includes:

Figures S1-10

Tables S1-2

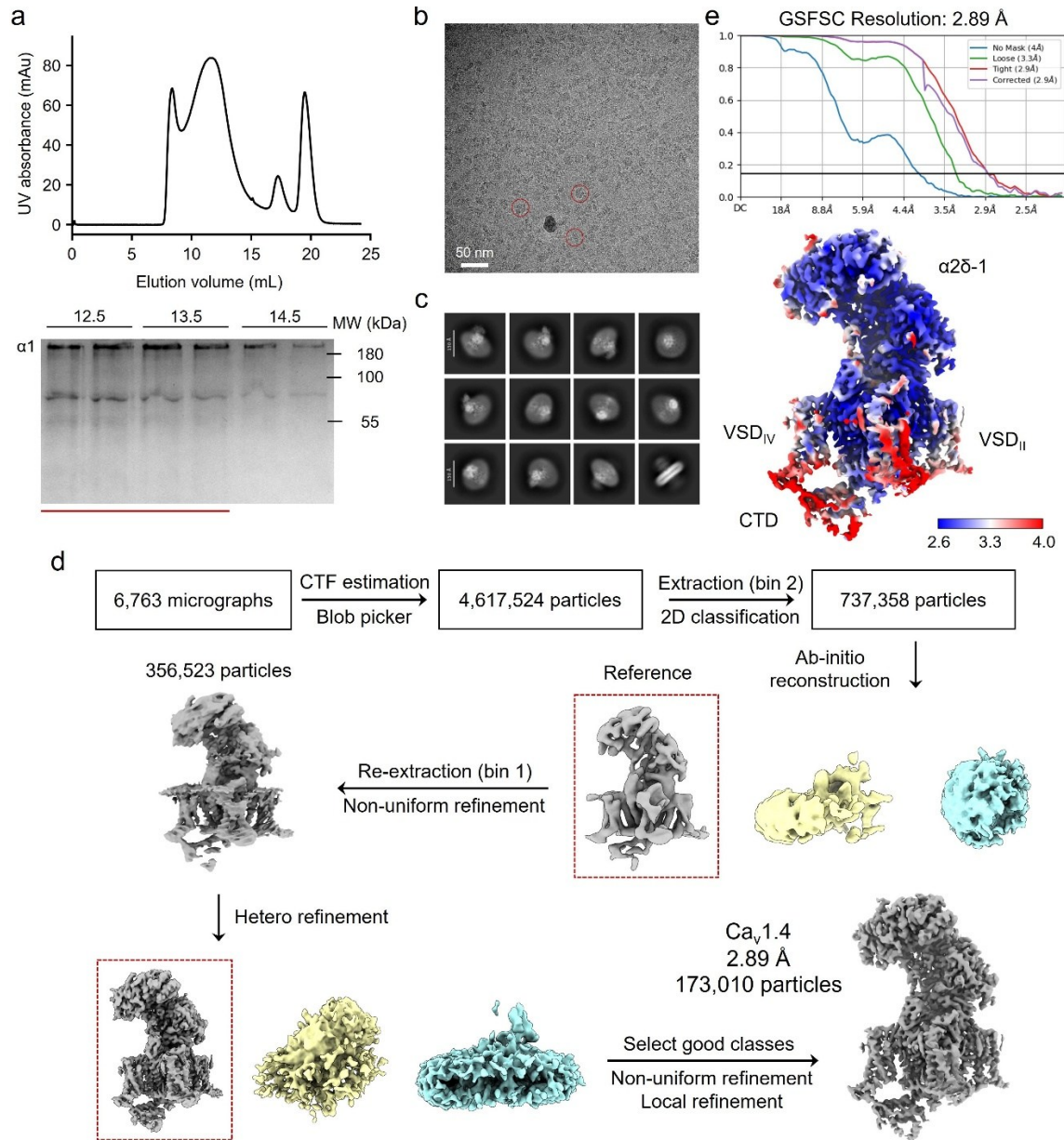


Figure S1 | Cryo-EM data analysis of the human $\text{Ca}_v1.4$ complex.

a, Size-exclusion chromatography and Coomassie blue staining analysis of the human $\text{Ca}_v1.4$ complex after two-step affinity purification. The indicated fractions were collected and concentrated for cryo-sample preparation. **b**, Representative cryo-EM micrograph of $\text{Ca}_v1.4$ complex. Particles with distinct views are indicated by red circles. Scale bar: 50 nm. **c**, Representative 2D class averages of $\text{Ca}_v1.4$ complex. Scale bar:

150 Å. **d**, Flowchart of cryo-EM data processing for Cav1.4 complex. Details can be found in Methods. **e**, Local resolution distribution and Golden Standard Fourier Shell Correlation (GSFSC) curve for the 3D reconstruction of Cav1.4 complex. All EM maps were contoured at 6 σ unless otherwise specified.

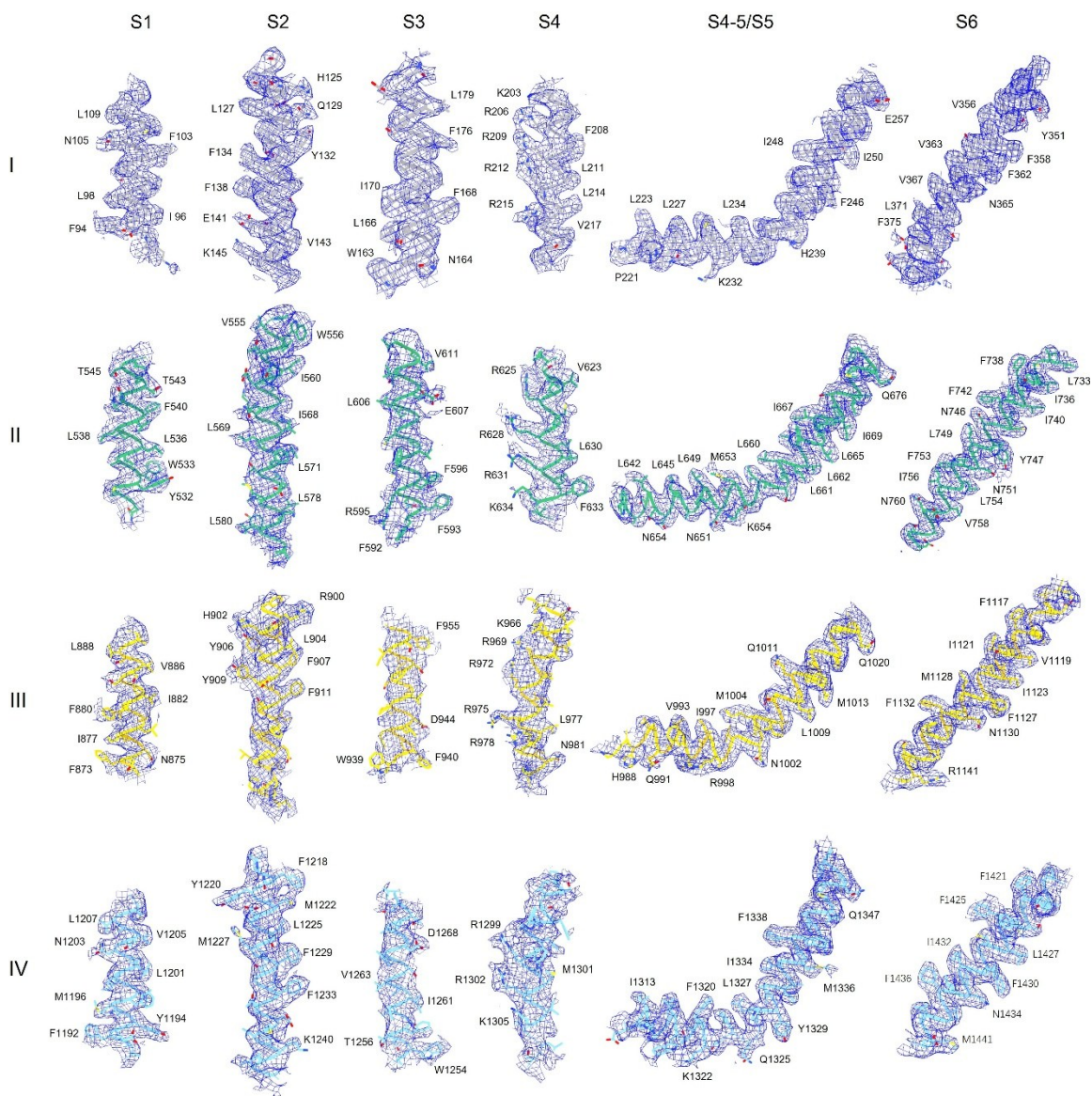


Figure S2 | EM maps for representative segments of $\text{Ca}_v1.4$.

EM maps for the transmembrane segments in each repeat of $\text{Ca}_v1.4$ $\alpha 1$ subunit. The densities are shown at a contour level of 4.5σ .

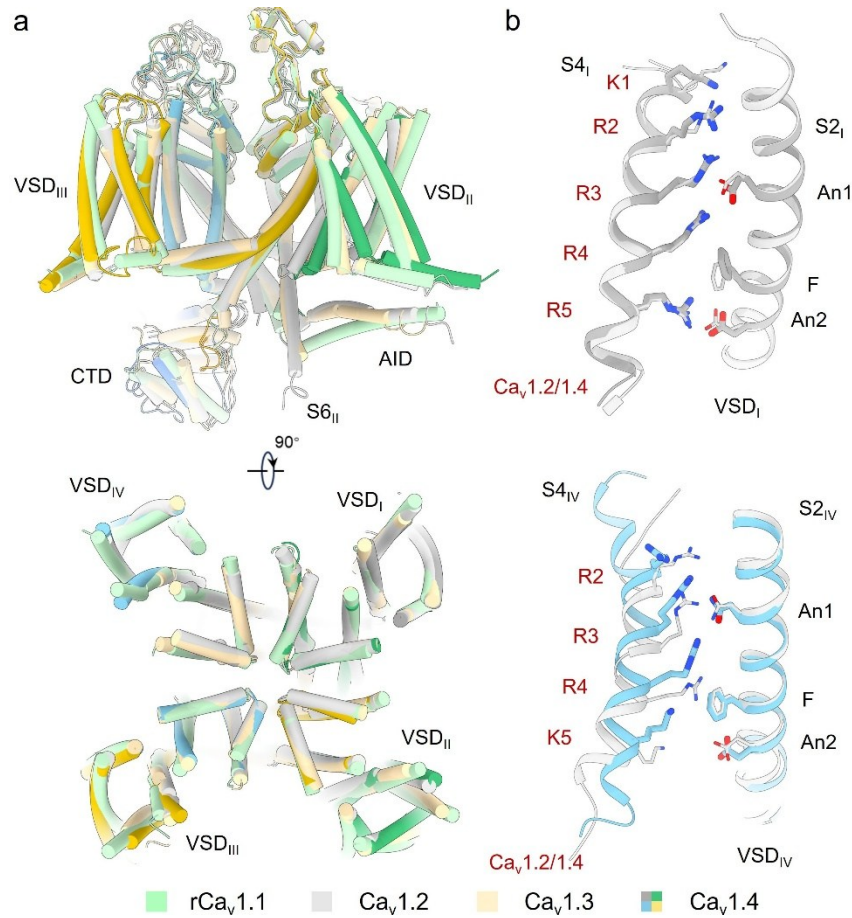


Figure S3 | Structural comparison of Cav1.4 with other reported Cav1 channels.

a, Two perpendicular views of superimposed structures are shown for rCav1.1 (light green, PDB: 5GJV), Cav1.2 (light grey, PDB: 8WE6), Cav1.3 (wheat, PDB: 7UHG), and Cav1.4 (domain-colored). The extracellular loops (ECLs) are omitted in the left panel for visual clarity. Major structural differences among these channels are observed in VSD_{II}.

b, The VSD_I and VSD_{IV} of Cav1.4 exhibit similar conformations to those of Cav1.2.

Structural superimposition of VSD_I and VSD_{IV} from Cav1.4 and Cav1.2 is shown. The S1 helices are omitted for visual clarity.

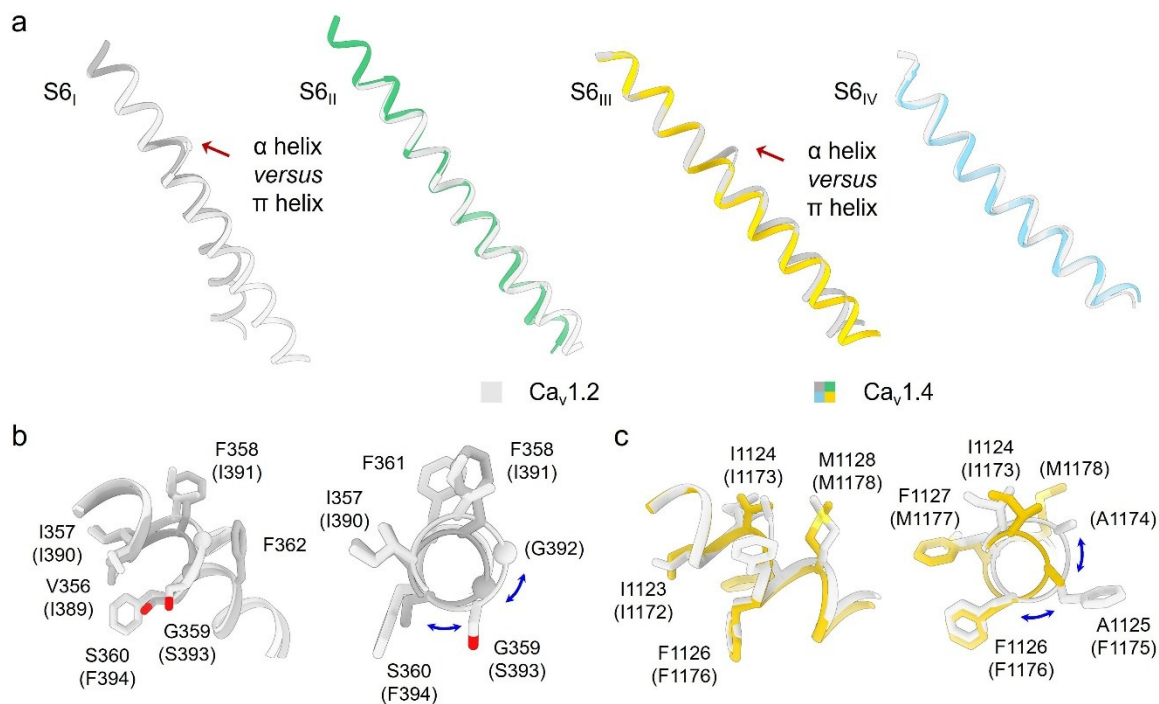


Figure S4 | Distinct structural conformers of S6_I and S6_{III} helices in Ca_v1.4.

a, Structural differences of S6 helices between Ca_v1.4 and Ca_v1.2. Superimposition of S6 helices reveals backbone differences between the α and π conformers. Both S6_I and S6_{III} of Ca_v1.4 adopt an α -helical conformation. **b**, **c**, Differences among α conformers in Ca_v1.4 arise from axial rotation of S6_I after G359 (**b**), and of S6_{III} after A1125 (**c**). The residues shown in brackets are from Ca_v1.2. Two views are shown.

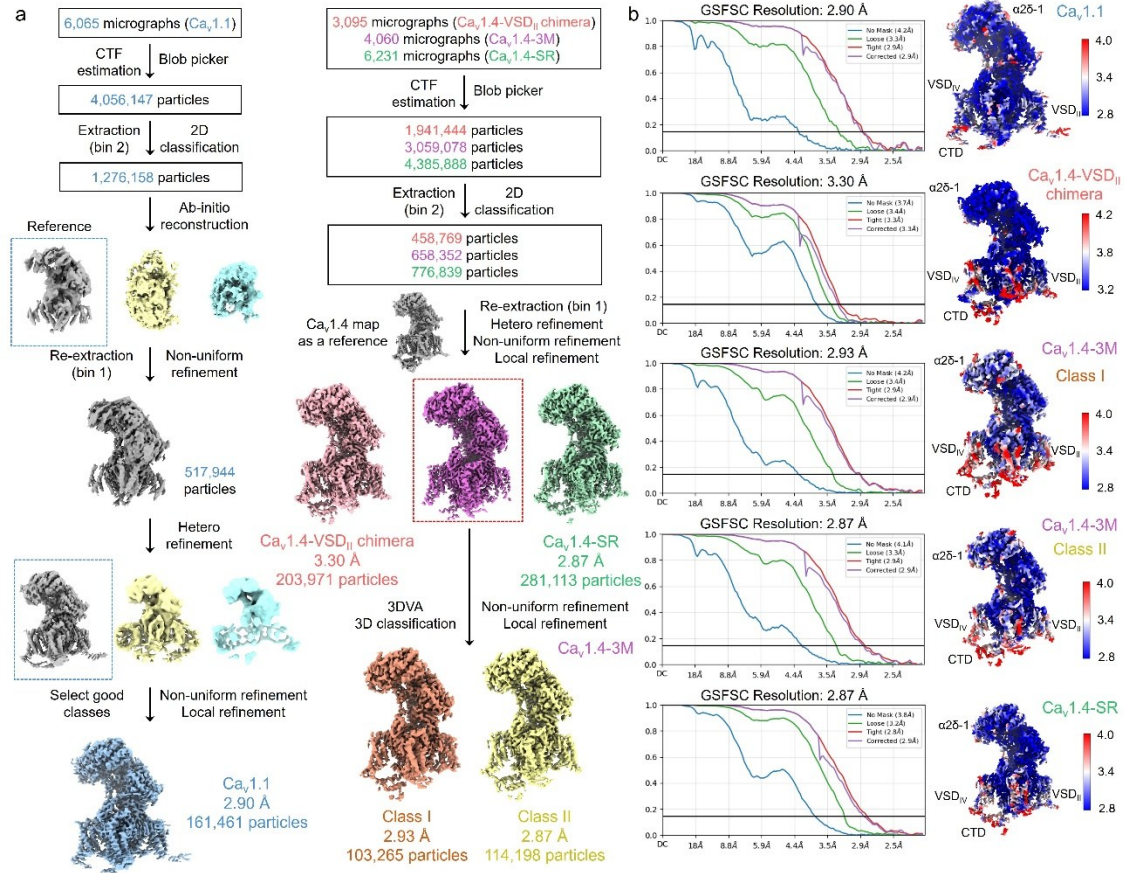


Figure S5 | Cryo-EM data analysis of the human Ca_v1.1 complex, Ca_v1.4 engineered variants, and Ca_v1.4 in complex with SR33805.

a, Flowcharts of cryo-EM data processing for human Ca_v1.1 complex, Ca_v1.4-VSD_{II}1.1 chimera, Ca_v1.4-3M, and Ca_v1.4-SR33805, respectively. Details can be found in Methods. **b**, Golden Standard Fourier Shell Correlation (GSFSC) curves and local resolution distributions for the 3D reconstructions of human Ca_v1.1 complex, Ca_v1.4-VSD_{II}1.1 chimera, Ca_v1.4-3M, and Ca_v1.4-SR33805, respectively.

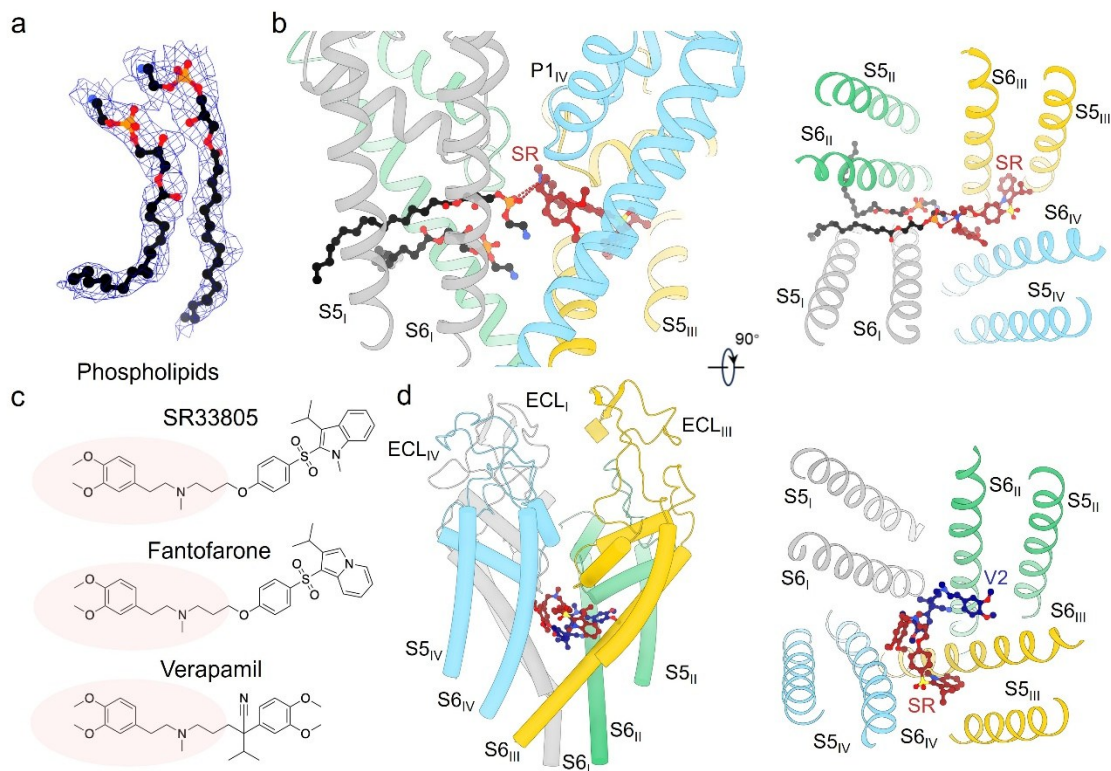


Figure S6 | Cryo-EM analysis of Cav1.4-SR33805.

a, Local EM density of the endogenous lipid. The EM map is contoured at 4 σ . **b**, The endogenous lipid stabilizes SR33805 binding in the central cavity of Cav1.4. Red dashed line indicates the potential H bond. **c**, Chemical structures of SR33805, fantofarone, and verapamil. The common chemical groups in these molecules are highlighted by the transparent pink oval. **d**, Structural comparison of SR33805 and verapamil (V2). Shown here are two perpendicular views. For clarity, only the PD of Cav1.4 is displayed. SR33805 and verapamil (V2) exhibit distinct binding poses, where SR33805 penetrates the fenestration formed by repeats III and IV, whereas verapamil (V2) is located near repeats II and III.

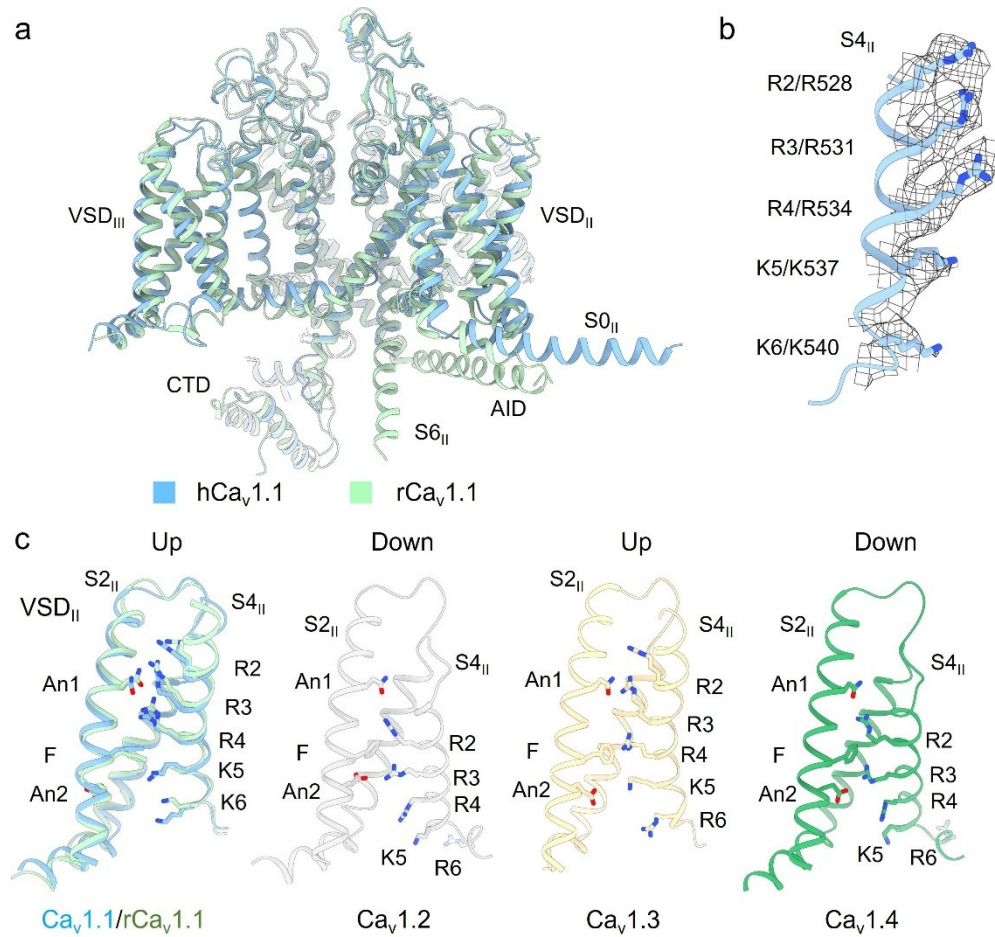


Figure S7 | Structural variations of VSD_{II} in Ca_v1 channels.

a, Structural comparison of human Ca_v1.1 and rabbit Ca_v1.1 reveals nearly identical conformations. **b**, Local EM density for S4_{II} of human Ca_v1.1. The EM density is contoured at 6 σ . **c**, Structural analysis indicates that the VSD_{II} of human Ca_v1.1, rabbit Ca_v1.1, and Ca_v1.3 exhibits an “up” conformation, whereas that of Ca_v1.2 and Ca_v1.4 adopts a “down” conformation. Human Ca_v1.1 is colored light blue, while the other Ca_v1 channels are colored according to the scheme used in Figure S3.

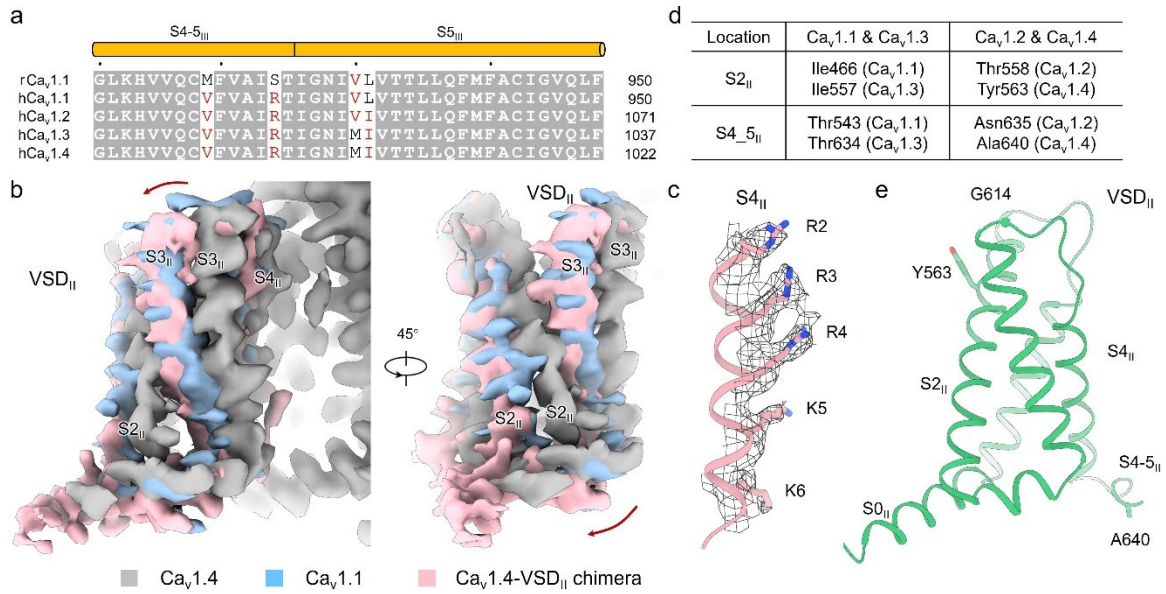


Figure S8 | Structural analysis of VSD_{II} in different conformations across Ca_v1 channels.

a, Sequence alignment of S4-5_{III} among Ca_v1 channels. **b**, Compared with Ca_v1.4, the VSD_{II} of Ca_v1.4-VSD_{II}1.1 chimera adopts a conformation resembling that of human Ca_v1.1. The EM maps for Ca_v1.4, human Ca_v1.1, and Ca_v1.4-VSD_{II}1.1 chimera are shown in grey, light blue, and pink, respectively. **c**, Local EM density for S4_{II} of Ca_v1.4-VSD_{II}1.1 chimera. The EM density is contoured at 6 σ . **d**, Sequence analysis identifies two residues that are conserved in Ca_v1.1 and Ca_v1.3, but variable in Ca_v1.2 and Ca_v1.4. **e**, Locations of the three mutated residues. Y563 is located on S2_{II}, G614 in S3-4 loop, and A640 in S4-5_{II}.

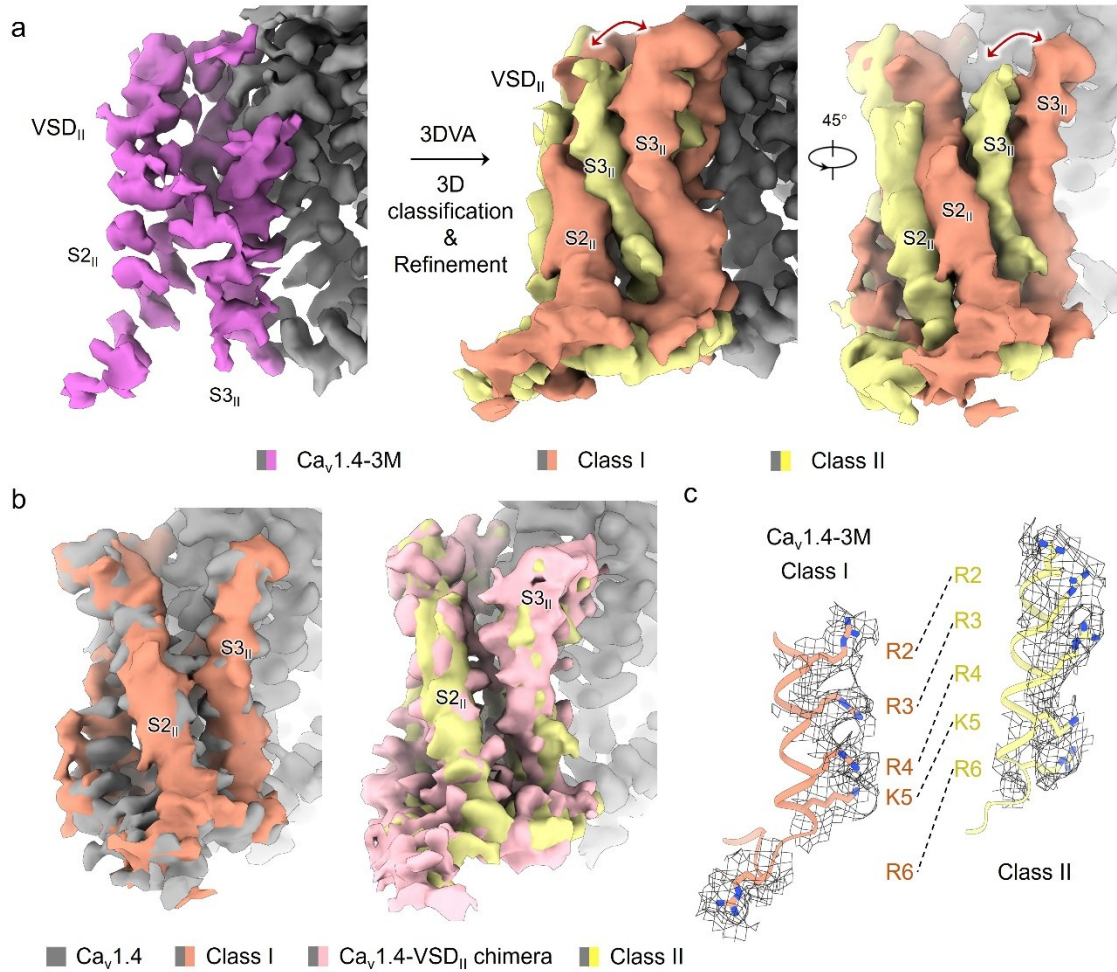


Figure S9 | Introducing triple mutations in $\text{Ca}_v1.4$ induces structural heterogeneity of VSD_{II} .

a, The poor local density for VSD_{II} in $\text{Ca}_v1.4\text{-3M}$ indicates increased structural dynamics of VSD_{II} after the introduction of triple mutations. Subsequent 3DVA, 3D classification, and refinement of $\text{Ca}_v1.4\text{-3M}$ generate two relatively stable conformations of VSD_{II} , termed Class I and Class II. The densities for VSD_{II} in $\text{Ca}_v1.4\text{-3M}$, Class I, and Class II are shown in magenta, orange, and yellow, respectively. Red arrows indicate the structural shifts. **b**, Structural comparison reveals that the conformation of VSD_{II} in Class I resembles that of WT $\text{Ca}_v1.4$, whereas that in Class II is similar to that of $\text{Ca}_v1.4\text{-VSD}_{II}1.1$ chimera. **c**, Local EM densities for S4_{II} of Class I and Class II. S4_{II} in Class I adopts a

“down” conformation, whereas that in Class II exhibits an “up” conformation. The EM density is contoured at 3.5σ .

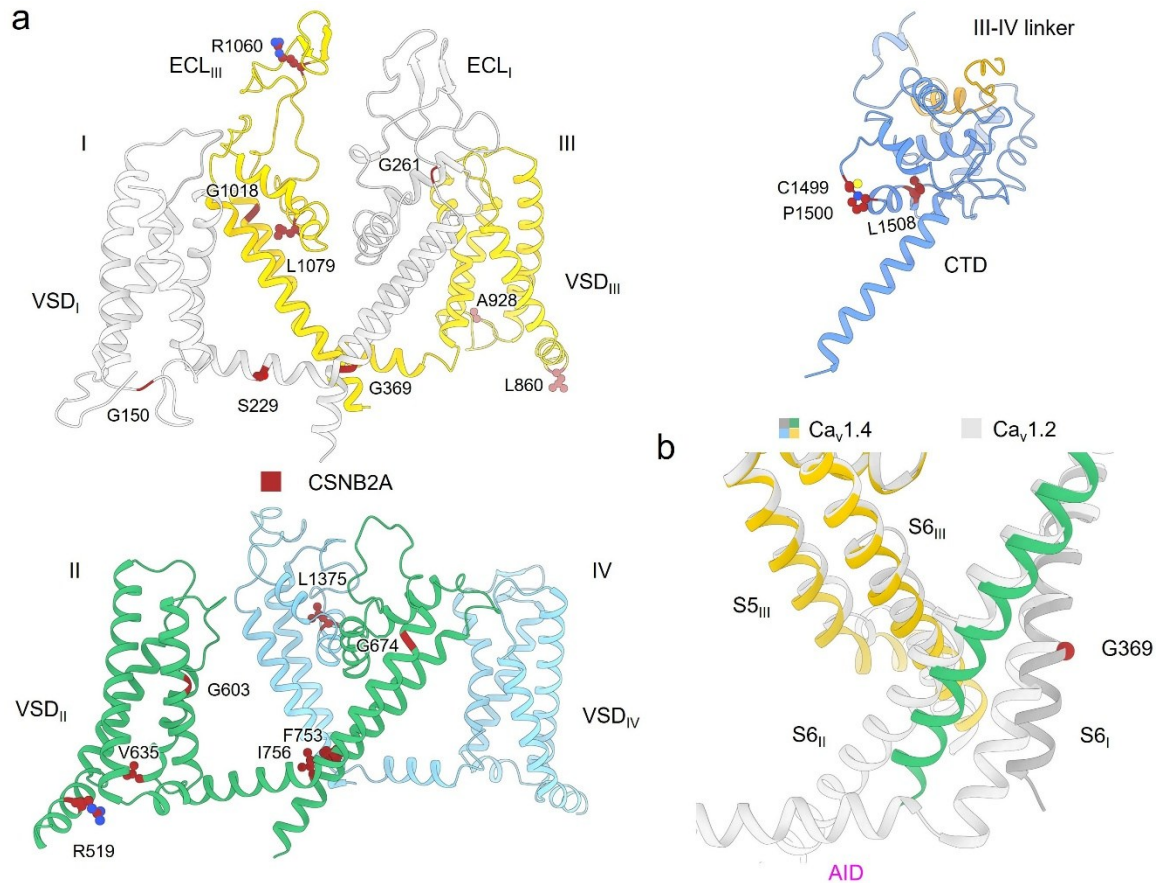


Figure S10 | Structural mapping of disease-associated mutations in Cav1.4.

a, Side views of repeat I and III, repeat II and IV, as well as the III-IV linker and CTD are shown. Please refer to Table S2 for details. CSNB2A: congenital stationary night blindness type 2A. **b**, G369, a disease-related mutation site, is located in S6_I and marks the starting point of the conformational shift of S6_I.

Table S1 | Cryo-EM data collection, refinement and validation statistics.

	Ca _v 1.4	Ca _v 1.1	Ca _v 1.4-VSD _{II} 1.1	Ca _v 1.4-3M Class I ClassII		Ca _v 1.4-SR33805
Data collection and processing						
Magnification	64,000	64,000	64,000	64,000		64,000
Voltage (kV)	300	300	300	300		300
Electron exposure (e ⁻ /Å ²)	50	50	50	50		50
Defocus range (μm)	-1.3 to -1.8	-1.3 to -1.8	-1.3 to -1.8	-1.3 to -1.8		-1.3 to -1.8
Pixel size (Å)	1.0979	1.0979	1.0979	1.0979		1.0979
Symmetry imposed	C1	C1	C1	C1		C1
Movies	6,763	6,065	3,095	4,060		6,231
Final particle images (No.)	173,010	161,461	203,971	103,265	114,198	281,113
Map resolution (Å)	2.89	2.90	3.30	2.93	2.87	2.87
FSC threshold	0.143	0.143	0.143	0.143	0.143	0.143
Refinement						
Map sharpening <i>B</i> factor (Å ²)	74.3	79.8	110.8	68.3	68.5	85.7
Model resolution (Å)	3.30	3.30	3.50	3.30	3.30	
FSC threshold	0.5	0.5	0.5	0.5	0.5	0.5
Model composition						
Non-hydrogen atoms	19,028	18,870	19,008	19,009	19,031	19,409
Protein residues	2,276	2,310	2,269	2,275	2,276	2,276
Ligands	38	23	38	38	38	39
<i>B</i> factors (Å ²)						
Protein	150.88	106.82	82.50	136.54	135.97	111.84
Ligand	157.29	123.79	101.42	136.64	142.85	104.86
R.m.s. deviations						
Bond lengths (Å)	0.004	0.005	0.004	0.003	0.003	0.003
Bond angles (°)	0.747	0.788	0.703	0.740	0.707	0.708
Validation						
MolProbity score	1.89	1.92	1.89	1.93	1.83	1.82
Clashscore	9.45	10.41	9.79	11.00	9.31	8.41
Poor rotamers (%)	0.65	0.49	0.30	0.25	0.60	0.30
Ramachandran plot						
Favored (%)	94.17	94.39	94.55	94.56	95.19	94.61
Allowed (%)	5.74	5.53	5.37	5.30	4.77	5.39
Outliers (%)	0.09	0.09	0.09	0.13	0.04	0.00

Table S2 | Disease-related mutations in Cav1.4.

Mutations	Disease	Location*	Mutations	Disease	Location
C74R	CSNB2A	NTD	I756T	CSNB2A	IS6II,35
G150R	CSNB2A	GS2I,39	L860P	CSNB2A	LS1III,9
S229P	CSNB2A	SS4I,50	A928D	CSNB2A	AS2III,40
G261R	CSNB2A	GS5I,45	G1018R	CSNB2A	GS5III,34
G369D	CSNB2A	GS6I,34	R1060W	CSNB2A	RS5III,87
R519Q	CSNB2A	RS1II,7	L1079P	CSNB2A	LP2III,21
G603R	CSNB2A	GS3II,36	L1375H	CSNB2A	LP2IV,20
V635I	CSNB2A	VS4II,37	C1499R	CSNB2A	CTD
G674D	CSNB2A	GS5II,34	P1500R	CSNB2A	CTD
G753C	CSNB2A	GS6II,32	L1508P	CSNB2A	CTD

CSNB2A: congenital stationary night blindness type 2

*The location information is based on the following reference:

Jin, X., Huang, J., Wang, H., Wang, K., & Yan, N. (2024). A versatile residue numbering scheme for Nav and Cav channels. *Cell Chemical Biology*, 31(8), 1394-1404.

Disease mutations in structurally unresolved regions are shaded light grey. Mutations are summarized from https://www.uniprot.org/uniprotkb/O60840/entry#disease_variants.